

Removing batch effects in gene expression array study

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Research Institutes

Data background (1)

- Hedenfalk et al. measured **3226** genes in seven BRCA1 and eight BRCA2 mutation-positive tumor samples.
- The goal of the study was to identify genes that showed differential expression across breast cancer tumor subtypes defined by these germline mutations.
- Several genes with apparent outliers were removed. This left **3170** genes.

Data background (2)

```
> SVA.pheno
```

```
      NEJM-PatientID Mutation
V7          1      BRCA1
V8          5      BRCA1
V9          3      BRCA1
V10         7      BRCA1
V11         2      BRCA1
V12         4      BRCA1
V13        10      BRCA2
V14         9      BRCA2
V15         8      BRCA2
V16        10      BRCA2
V17         6      BRCA1
V18        13      BRCA2
V19        14      BRCA2
V20        11      BRCA2
V21        12      BRCA2
```

```
> head(SVA.exp.preprocessed)
```

```
      q-value      p-value fold-change (log base 2) s1996 s1822 s1714 s1224 s1252 s1510 s1900 s1787 s1721 s1486 s1905
[1,] 0.089529 0.0122344      1.203 0.15 0.22 0.30 0.26 1.22 0.44 0.35 1.10 1.07 1.46 0.38
[2,] 0.213752 0.0761198     -0.521 1.54 1.27 0.76 0.85 1.27 0.64 0.90 0.64 0.78 0.55 0.61
[3,] 0.672438 0.99530284      0.002 1.72 1.57 2.13 1.09 1.98 0.74 1.71 1.16 1.33 1.46 2.43
[4,] 0.163987 0.04212934      0.690 0.71 1.24 1.69 2.23 1.16 0.82 1.44 2.03 3.60 1.20 2.08
[5,] 0.637670 0.84745741      0.053 0.94 1.53 1.87 1.19 1.16 1.54 1.05 0.91 0.85 1.22 1.01
[6,] 0.377451 0.25437224     -0.227 0.80 0.95 1.53 1.37 1.02 1.22 0.78 0.96 0.65 1.02 1.09

      s1816 s1616 s1063 s1936
[1,] 0.73 0.63 0.77 0.66
[2,] 0.71 0.30 0.62 1.00
[3,] 1.71 1.26 1.41 3.00
[4,] 3.24 2.41 1.56 2.56
[5,] 3.25 2.20 1.09 1.29
[6,] 0.66 1.40 1.32 1.13
```

```
> head(exp.annotation)
```

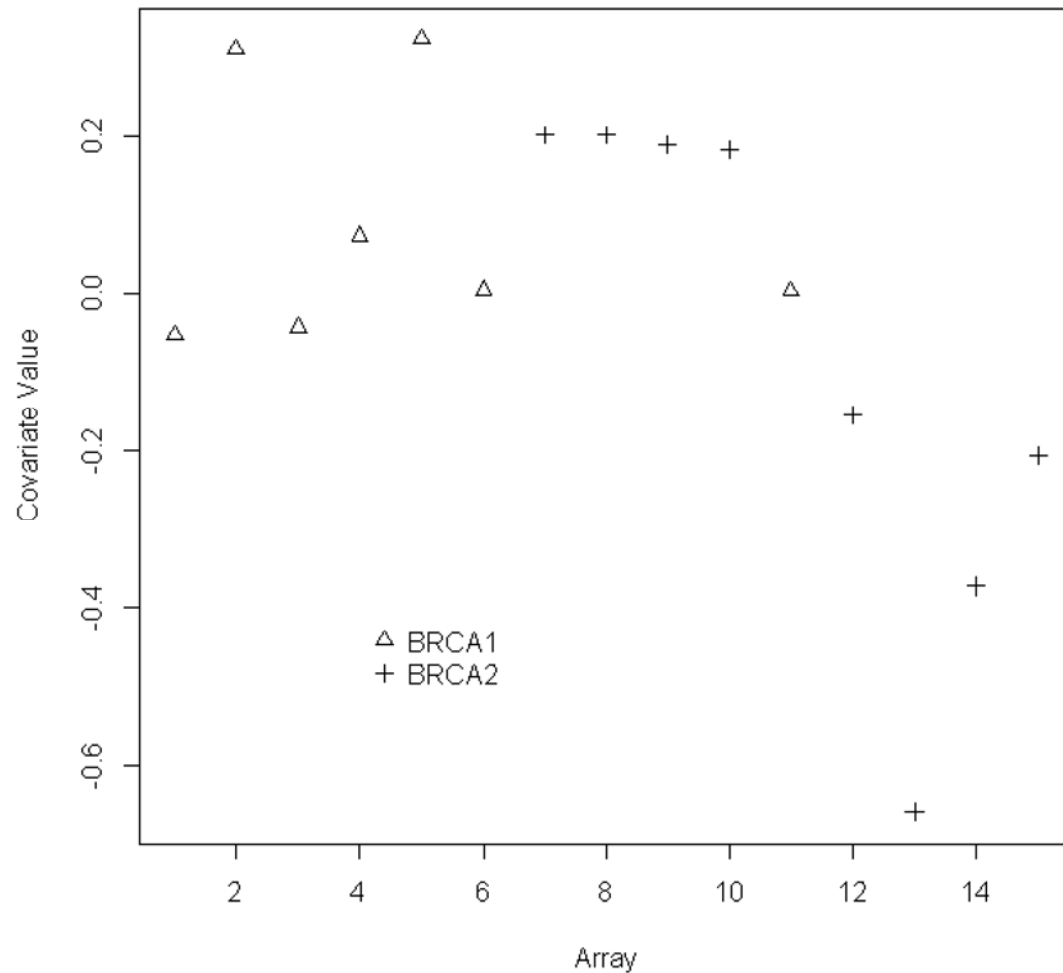
```
      PlatePosition ImageCloneID
4          HK1A1      21652
5          HK1A2      22012
6          HK1A4      22293
7          HK1A5      22493
8          HK1A6      23019
9          HK1A7      23132

      Title
catenin (cadherin-associated protein), alpha 1 (102kD)
ADP-ribosylation factor 3
uroporphyrinogen III synthase (congenital erythropoietic porphyria)
ribosomal protein L26
guanine nucleotide binding protein (G protein), alpha stimulating activity polypeptide 1
pre-mRNA splicing factor SF3a (120 kDa subunit), similar to S. cerevisiae PRP21
```

Substructure detected

- Hierarchical clustering of the data reveals notable substructure within the BRCA2 samples.
- Applied **SVA** (Surrogate Variable Analysis) to identified a single surrogate variable that appears to capture this trend.

Substructure detected



Significance analysis

- Included this surrogate variable in a significance analysis comparing BRCA1 and BRCA2 tumors.
- The number of genes differentially expressed between BRCA1 and BRCA2 before and after adjusting for surrogate variables.

Analysis Type	q-Value Threshold			
	0.01	0.025	0.05	0.10
Unadjusted	1	19	96	275
SVA adjusted	0	10	48	190

R package: sva (1)

- Create the full model matrix - including both the adjustment variables and the variable of interest (BRCA1/BRCA2).
- The null model contains only the adjustment variables.

```
> mod=model.matrix(~as.factor(Mutation),data=SVB.pheno)
> mod0=model.matrix(~1,data=SVB.pheno)
> svaObj=sva(log2(SVB.exp.preprocessed[,-1:-3]),mod,mod0)
Number of significant surrogate variables is: 4
Iteration (out of 5 ):1 2 3 4 5 > |
```

R package: sva (2)

- The `f.pvalue` function can be used to calculate parametric F-test p-values for each gene. The F-test compares the models `mod` and `mod0`.

```
> pValues.before=f.pvalue(log2(SVA.exp.preprocessed[,-1:-3]),mod,mod0)
> qValuesObj.before<-qvalue(pValues.before)
> qsummary(qValuesObj.before)
```

Call:

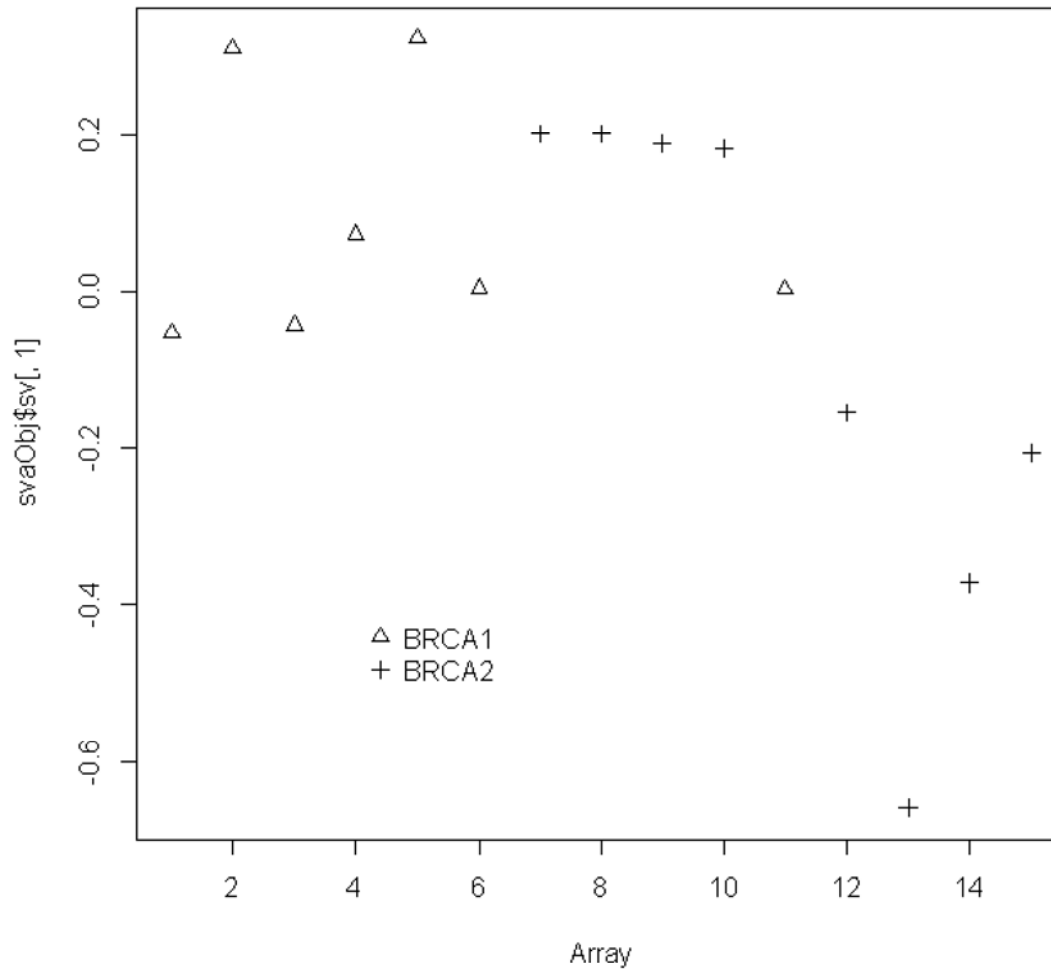
```
qvalue(p = pValues.before)
```

pi0: 0.6786508

Cumulative number of significant calls:

	<1e-04	<0.001	<0.01	<0.025	<0.05	<0.1	<1
p-value	9	62	228	392	565	832	3170
q-value	0	0	1	19	96	275	3170

R package: sva (3)



R package: sva (4)

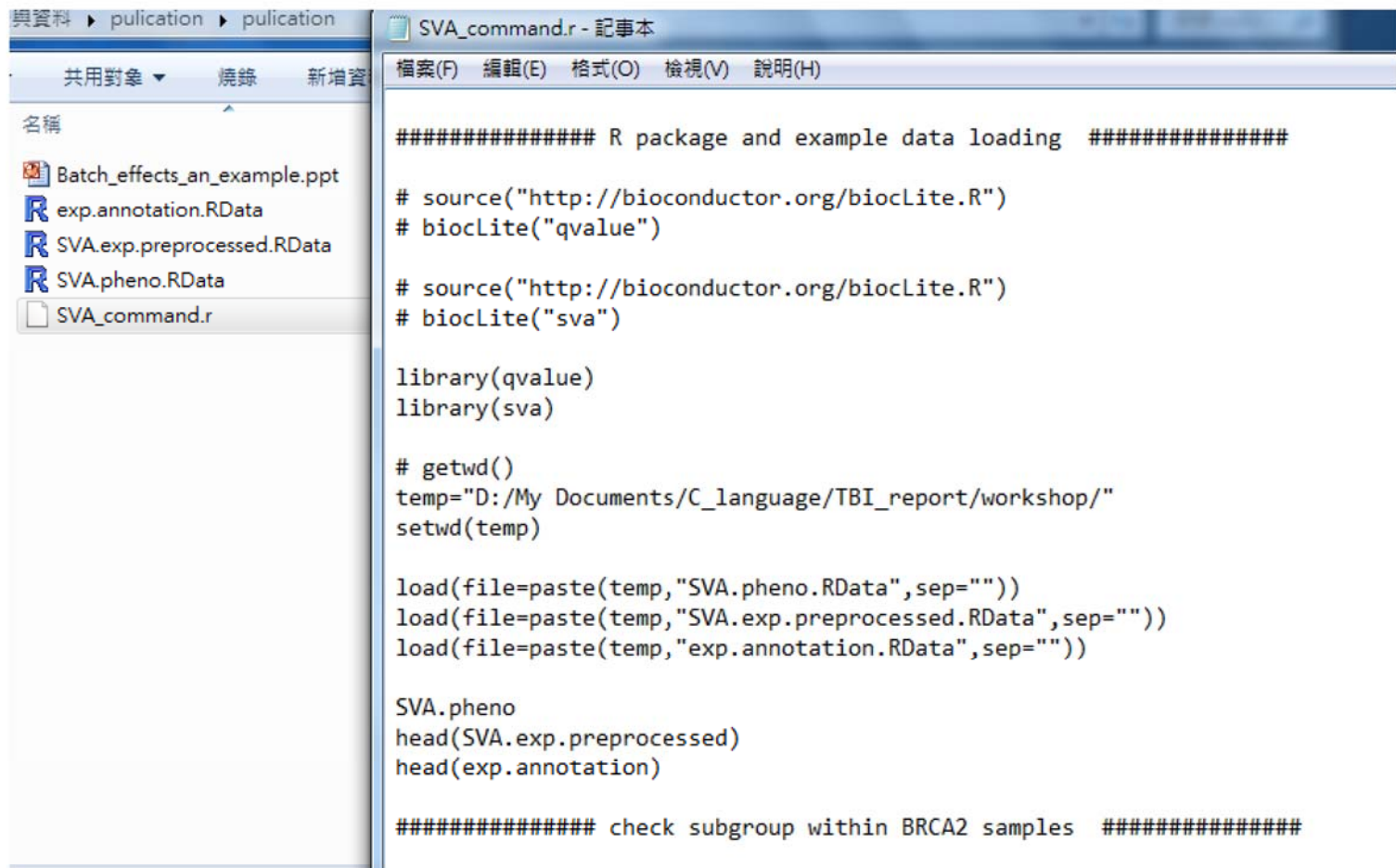
- Now we can perform the same analysis, but adjusting for surrogate variables. The first step is to include the surrogate variables in both the null and full models.

```
> svaObj=sva(log2(SVA.exp.preprocessed[,-1:-3]),mod,mod0)
Number of significant surrogate variables is: 4
Iteration (out of 5 ):1 2 3 4 5 >
> modSv=cbind(mod,svaObj$sv[,1])
> mod0Sv=cbind(mod0,svaObj$sv[,1])
```

- Then P-values and Q-values can be computed as before.

Step by Step: Open the text file

Use the Notepad to open the file named "SVA_command.r" (text file).



The screenshot shows a Windows file explorer window on the left and a Notepad window on the right. The file explorer window displays a folder named 'pulation' containing several files, including 'SVA_command.r'. The Notepad window, titled 'SVA_command.r - 記事本', contains R code for loading data and performing analysis.

```
##### R package and example data loading #####  
  
# source("http://bioconductor.org/biocLite.R")  
# biocLite("qvalue")  
  
# source("http://bioconductor.org/biocLite.R")  
# biocLite("sva")  
  
library(qvalue)  
library(sva)  
  
# getwd()  
temp="D:/My Documents/C_language/TBI_report/workshop/"  
setwd(temp)  
  
load(file=paste(temp,"SVA.pheno.RData",sep=""))  
load(file=paste(temp,"SVA.exp.preprocessed.RData",sep=""))  
load(file=paste(temp,"exp.annotation.RData",sep=""))  
  
SVA.pheno  
head(SVA.exp.preprocessed)  
head(exp.annotation)  
  
##### check subgroup within BRCA2 samples #####
```

Step by Step: Install R packages (1)

```
##### R package and example data loading #####
```

```
source("http://bioconductor.org/biocLite.R")
biocLite("qvalue")
```

1. Copy the command lines for installing the R package "qvalue".

```
source("http://bioconductor.org/biocLite.R")
biocLite("qvalue")

library(qvalue)
library(qvalue)

# get the example data
temp = tempfile()
setwd(temp)

load("data", se
load("data", se
load("data", se
load("data", se

SVA.pheno
head(SVA.exp.preprocessed)
head(exp.annotation)
```

復原(U)
剪下(T)
複製(C)
貼上(P)
刪除(D)
全選(A)
從右到左的讀取順序(R)
顯示 Unicode 控制字元(S)
插入 Unicode 控制字元(I)
開啟 IME(O)
重新轉換(R)

R GUI (32-bit)

File Edit View Misc Packages Windows Help

R Console

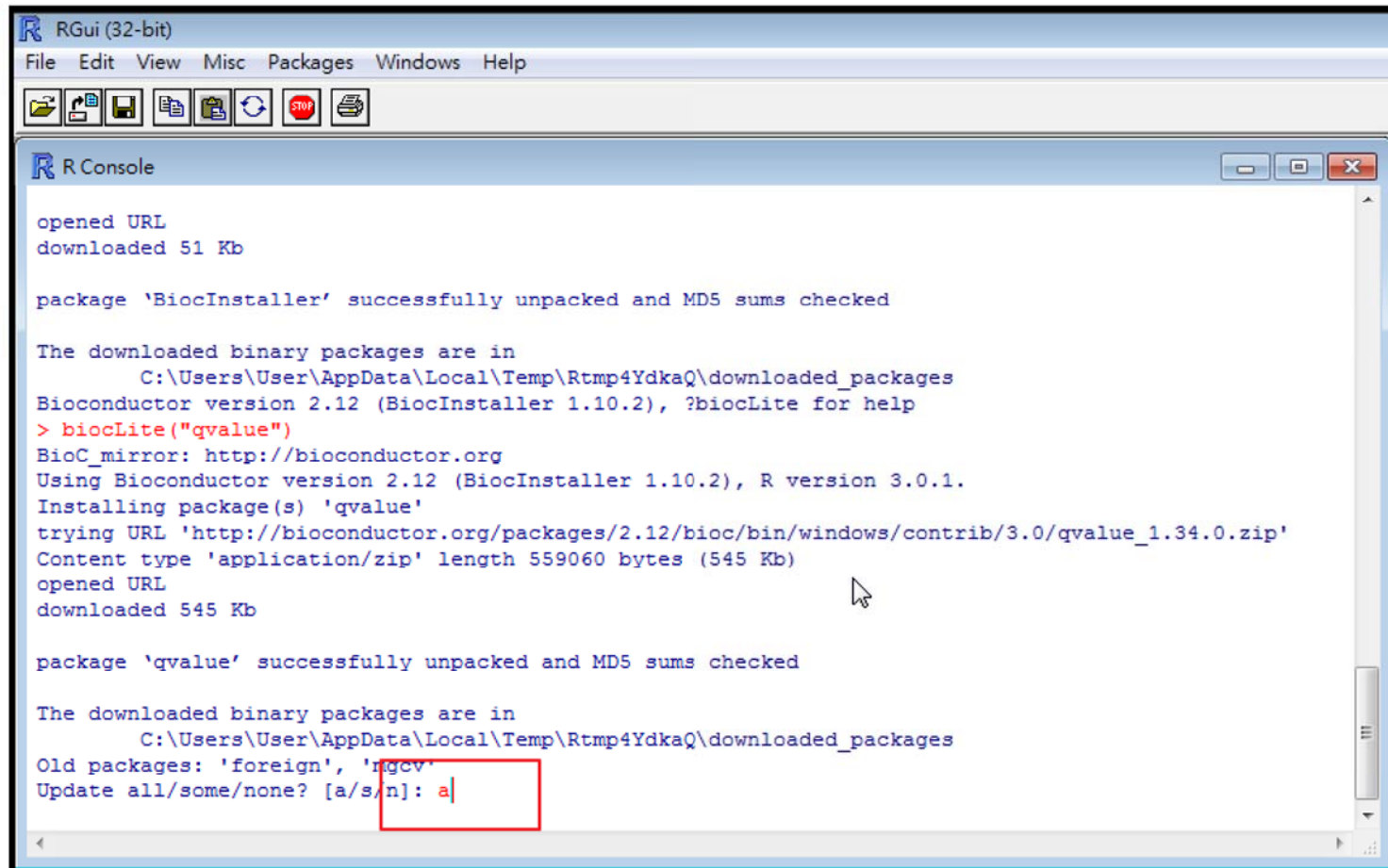
```
[1,] 1.22 0.44 0.35 1.10 1.07 1.46 0.38 0.73 0.63 0.77 0.66
[2,] 1.27 0.64 0.90 0.64 0.78 0.55 0.61 0.71 0.30 0.62 1.00
[3,] 1.98 0.74 1.71 1.16 1.33 1.46 2.43 1.71 1.26 1.41 3.00
[4,] 1.16 0.82 1.44 2.03 3.60 1.20 2.08 3.24 2.41 1.56 2.56
[5,] 1.16 1.54 1.05 0.91 0.85 1.22 1.01 3.25 2.20 1.09 1.29
[6,] 1.02 1.22 0.78 0.96 0.65 1.02 1.09 0.66 1.40 1.32 1.13

> head(exp.annotation)
  PlatePosition ImageCloneID
4          HK1A1       21652
5          HK1A2       22012
6          HK1A4       22293
7          HK1A5       22493
8          HK1A6       23019
9          HK1A7       23132

catenin (cadherin-associated protein), alpha 1 (102kD)
ADP-ribosylation factor 3
uroporphyrinogen III synthase (congenital erythropoietic porphyria)
ribosomal protein L26
guanine nucleotide binding protein (G protein), alpha stimulating activity polypeptide 1
pro-mRNA splicing factor SFPQ (120 kDa subunit), similar to S. cerevisiae PBP21
```

2. Paste to the R window.

Step by Step: Install R packages (2)



```
RGui (32-bit)
File Edit View Misc Packages Windows Help

R Console

opened URL
downloaded 51 Kb

package 'BiocInstaller' successfully unpacked and MD5 sums checked

The downloaded binary packages are in
  C:\Users\User\AppData\Local\Temp\Rtmp4YdkaQ\downloaded_packages
Bioconductor version 2.12 (BiocInstaller 1.10.2), ?biocLite for help
> biocLite("qvalue")
BioC_mirror: http://bioconductor.org
Using Bioconductor version 2.12 (BiocInstaller 1.10.2), R version 3.0.1.
Installing package(s) 'qvalue'
trying URL 'http://bioconductor.org/packages/2.12/bioc/bin/windows/contrib/3.0/qvalue_1.34.0.zip'
Content type 'application/zip' length 559060 bytes (545 Kb)
opened URL
downloaded 545 Kb

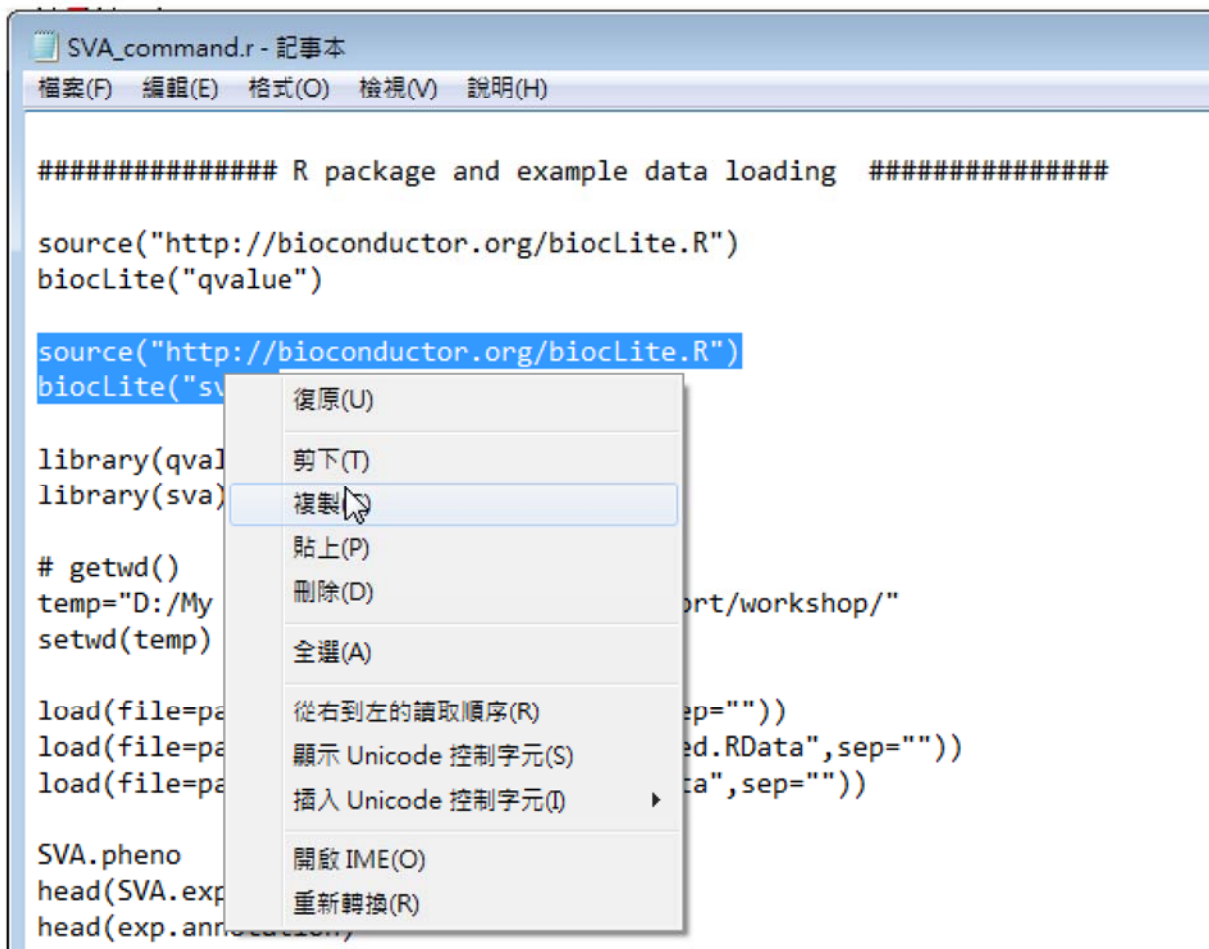
package 'qvalue' successfully unpacked and MD5 sums checked

The downloaded binary packages are in
  C:\Users\User\AppData\Local\Temp\Rtmp4YdkaQ\downloaded_packages
Old packages: 'foreign', 'mgcv'
Update all/some/none? [a/s/n]: a
```

3. Type “a” for updating all old packages.

Step by Step: Install R packages (3)

4. Repeat step 1-3. The same way to install the package "sva".



```
SVA_command.r - 記事本
檔案(F)  編輯(E)  格式(O)  檢視(V)  說明(H)

##### R package and example data loading #####

source("http://bioconductor.org/biocLite.R")
biocLite("qvalue")

source("http://bioconductor.org/biocLite.R")
biocLite("sva")

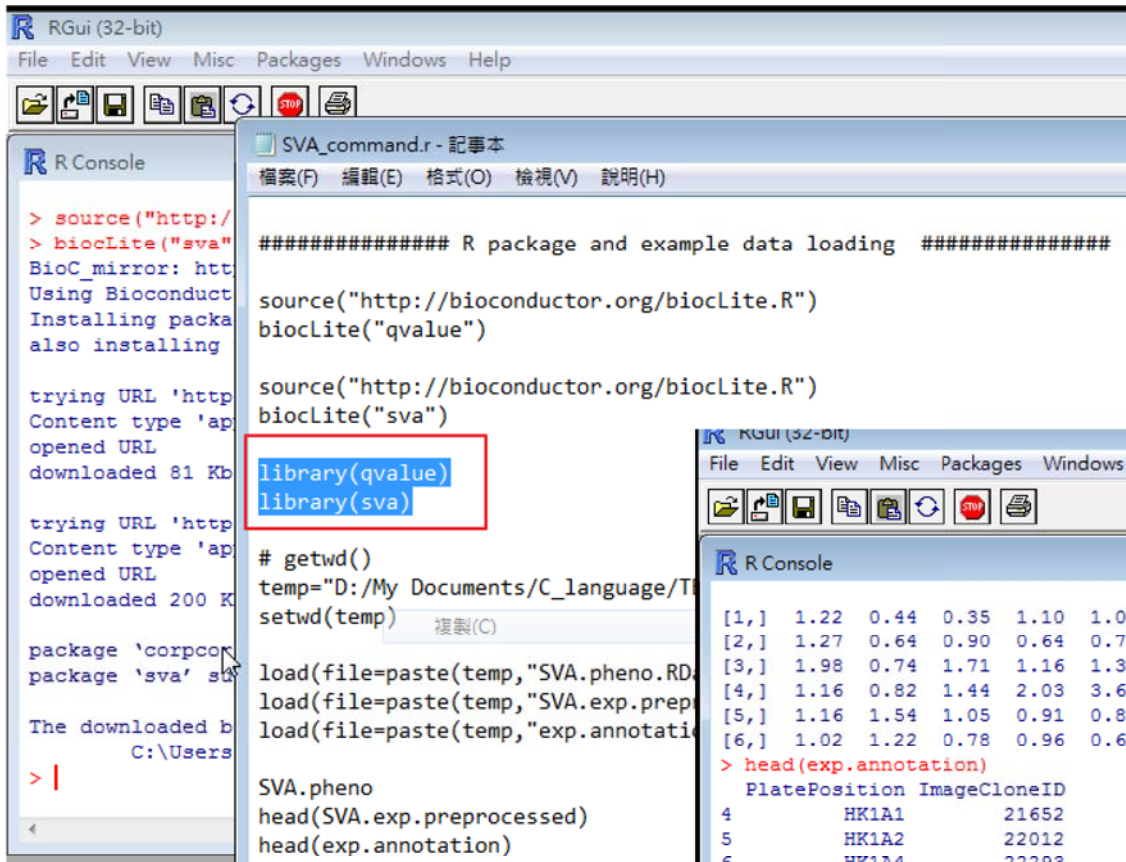
library(qvalue)
library(sva)

# getwd()
temp="D:/My
setwd(temp)

load(file=paste0(temp, "data", sep=""))
load(file=paste0(temp, "data", sep=""))
load(file=paste0(temp, "data", sep=""))

SVA.pheno
head(SVA.exp)
head(exp.anno)
```

Step by Step : Load R packages



The screenshot shows the RGui (32-bit) interface. The R Console on the left displays the output of the commands entered in the Notepad window. The Notepad window, titled 'SVA_command.r - 記事本', contains the following R code:

```
##### R package and example data loading #####

source("http://bioconductor.org/biocLite.R")
biocLite("qvalue")

source("http://bioconductor.org/biocLite.R")
biocLite("sva")

library(qvalue)
library(sva)

# getwd()
temp="D:/My Documents/C_language/T
setwd(temp)

load(file=paste(temp,"SVA.pheno.RD
load(file=paste(temp,"SVA.exp.prep
load(file=paste(temp,"exp.annotati

SVA.pheno
head(SVA.exp.preprocessed)
head(exp.annotation)
```

The R Console shows the following output:

```
> source("http://
> biocLite("sva"
BioC_mirror: htt
Using Bioconduct
Installing packa
also installing

trying URL 'http
Content type 'ap
opened URL
downloaded 81 Kb

trying URL 'http
Content type 'ap
opened URL
downloaded 200 K

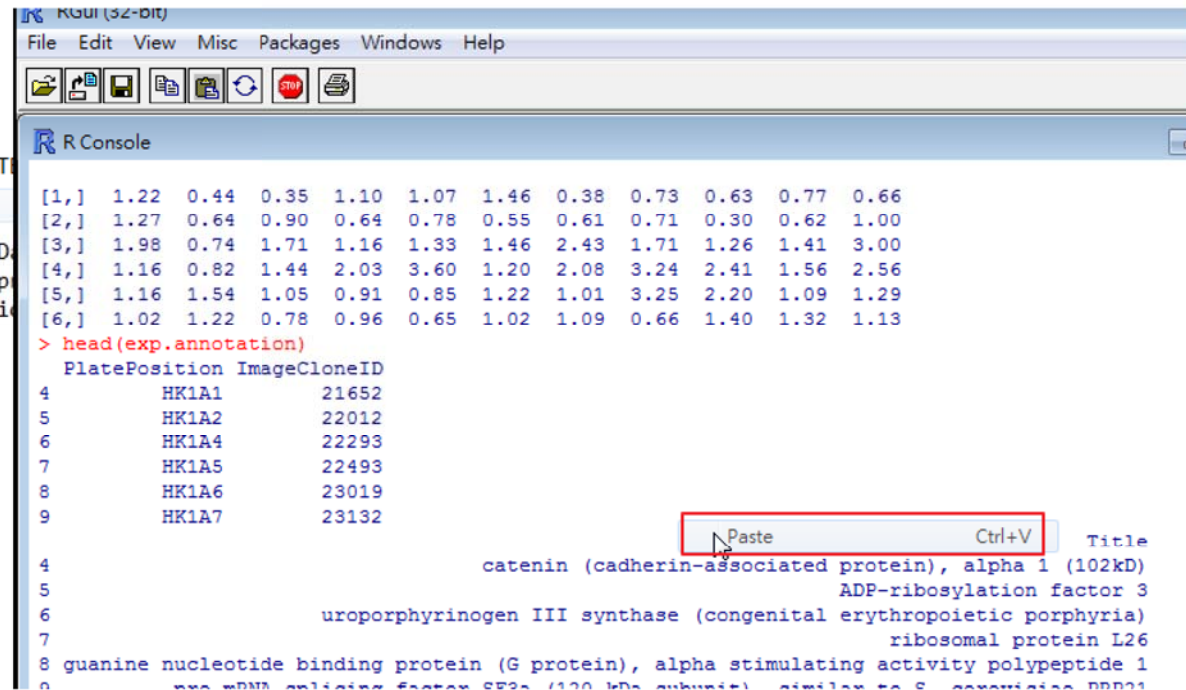
package 'corpcor
package 'sva' sta

The downloaded b
C:\Users

> |
```

1. Copy the command lines for loading two R packages, "qvalue" and "sva".

2. Paste to the R window.



The screenshot shows the RGui (32-bit) interface. The R Console on the right displays the output of the commands entered in the Notepad window. The Notepad window, titled 'SVA_command.r - 記事本', contains the following R code:

```
##### R package and example data loading #####

source("http://bioconductor.org/biocLite.R")
biocLite("qvalue")

source("http://bioconductor.org/biocLite.R")
biocLite("sva")

library(qvalue)
library(sva)

# getwd()
temp="D:/My Documents/C_language/T
setwd(temp)

load(file=paste(temp,"SVA.pheno.RD
load(file=paste(temp,"SVA.exp.prep
load(file=paste(temp,"exp.annotati

SVA.pheno
head(SVA.exp.preprocessed)
head(exp.annotation)
```

The R Console shows the following output:

```
[1,] 1.22 0.44 0.35 1.10 1.07 1.46 0.38 0.73 0.63 0.77 0.66
[2,] 1.27 0.64 0.90 0.64 0.78 0.55 0.61 0.71 0.30 0.62 1.00
[3,] 1.98 0.74 1.71 1.16 1.33 1.46 2.43 1.71 1.26 1.41 3.00
[4,] 1.16 0.82 1.44 2.03 3.60 1.20 2.08 3.24 2.41 1.56 2.56
[5,] 1.16 1.54 1.05 0.91 0.85 1.22 1.01 3.25 2.20 1.09 1.29
[6,] 1.02 1.22 0.78 0.96 0.65 1.02 1.09 0.66 1.40 1.32 1.13

> head(exp.annotation)
  PlatePosition ImageCloneID
4          HK1A1         21652
5          HK1A2         22012
6          HK1A4         22293
7          HK1A5         22493
8          HK1A6         23019
9          HK1A7         23132

catenin (cadherin-associated protein), alpha 1 (102kD)
ADP-ribosylation factor 3
uroporphyrinogen III synthase (congenital erythropoietic porphyria)
ribosomal protein L26
guanine nucleotide binding protein (G protein), alpha stimulating activity polypeptide 1
xxx mRNA encoding factor SFPs (120 kDa subunit) similar to S. cerevisiae RRP21
```

Step by Step : R environment

1. Change the current working directory for loading the example data files.
2. Replace the symbol “\” to “/”.



```
SVA_command.r - 記事本
檔案(F) 編輯(E) 格式(O) 檢視(V) 說明(H)

source("http://bioconductor.org/biocLite.R")
biocLite("qvalue")

source("http://bioconductor.org/biocLite.R")
biocLite("sva")

library(qvalue)
library(sva)

# getwd()
temp="C:\\Users\\User\\Desktop\\請安裝軟體與資料\\publication\\publication/"
setwd(temp)

load(file=paste(temp, "SVA.pheno.RData", sep=""))
load(file=paste(temp, "SVA.exp.preprocessed.RData", sep=""))
load(file=paste(temp, "exp.annotation.RData", sep=""))

SVA.pheno
head(SVA.exp.preprocessed)
head(exp.annotation)
```

Step by Step : Load example files

```
SVA_command.r - 記事本
檔案(F) 編輯(E) 格式(O) 檢視(V) 說明(H)

source("http://bioconductor.org/biocLite.R")
biocLite("qvalue")

source("http://bioconductor.org/biocLite.R")
biocLite("sva")

library(qvalue)
library(sva)

# getwd()
temp="C:/Users/User/Desktop/請安裝軟體
setwd(temp)

load(file=paste(temp, "SVA.pheno.RData")
load(file=
load(file=
SVA.pheno
head(SVA.
head(exp.
```

1. Copy the command lines for loading example files.

2. Paste to the R window.

```
R GUI (32-bit)
File Edit View Misc Packages Windows Help

R Console

[1,] 1.22 0.44 0.35 1.10 1.07 1.46 0.38 0.73 0.63 0.77 0.66
[2,] 1.27 0.64 0.90 0.64 0.78 0.55 0.61 0.71 0.30 0.62 1.00
[3,] 1.98 0.74 1.71 1.16 1.33 1.46 2.43 1.71 1.26 1.41 3.00
[4,] 1.16 0.82 1.44 2.03 3.60 1.20 2.08 3.24 2.41 1.56 2.56
[5,] 1.16 1.54 1.05 0.91 0.85 1.22 1.01 3.25 2.20 1.09 1.29
[6,] 1.02 1.22 0.78 0.96 0.65 1.02 1.09 0.66 1.40 1.32 1.13
> head(exp.annotation)
  PlatePosition ImageCloneID
4          HK1A1       21652
5          HK1A2       22012
6          HK1A4       22293
7          HK1A5       22493
8          HK1A6       23019
9          HK1A7       23132

4          catenin (cadherin-associated protein), alpha 1 (102kD)
5                      ADP-ribosylation factor 3
6          uroporphyrinogen III synthase (congenital erythropoietic porphyria)
7                      ribosomal protein L26
8 guanine nucleotide binding protein (G protein), alpha stimulating activity polypeptide 1
9          xxx mRNA, partial, clone SF25 (120 kDa subunit), similar to S. cerevisiae PBP21
```

Paste Ctrl+V

Step by Step : Command lines

The package “sva” command lines.

```
mod=model.matrix(~as.factor(Mutation),data=SVA.pheno)
mod0=model.matrix(~1,data=SVA.pheno)
svaObj=sva(log2(SVA.exp.preprocessed[,-1:-3]),mod,mod0)
```

The command for plotting the graph.

```
symbol<-rep(2,15)
symbol[BRCA2]<-3
svaObj$sv

plot(1:15,svaObj$sv[,1],xlab="Array",ylab="Covariate Value",pch=symbol)
legend(4,-0.4,c("BRCA1","BRCA2"),pch=c(2,3),bty = "n")
```

The command for computing F-test and q-value.

```
##### compare q-value before/after batch effect #####

pValues.before=f.pvalue(log2(SVA.exp.preprocessed[,-1:-3]),mod,mod0)
qValuesObj.before<-qvalue(pValues.before)

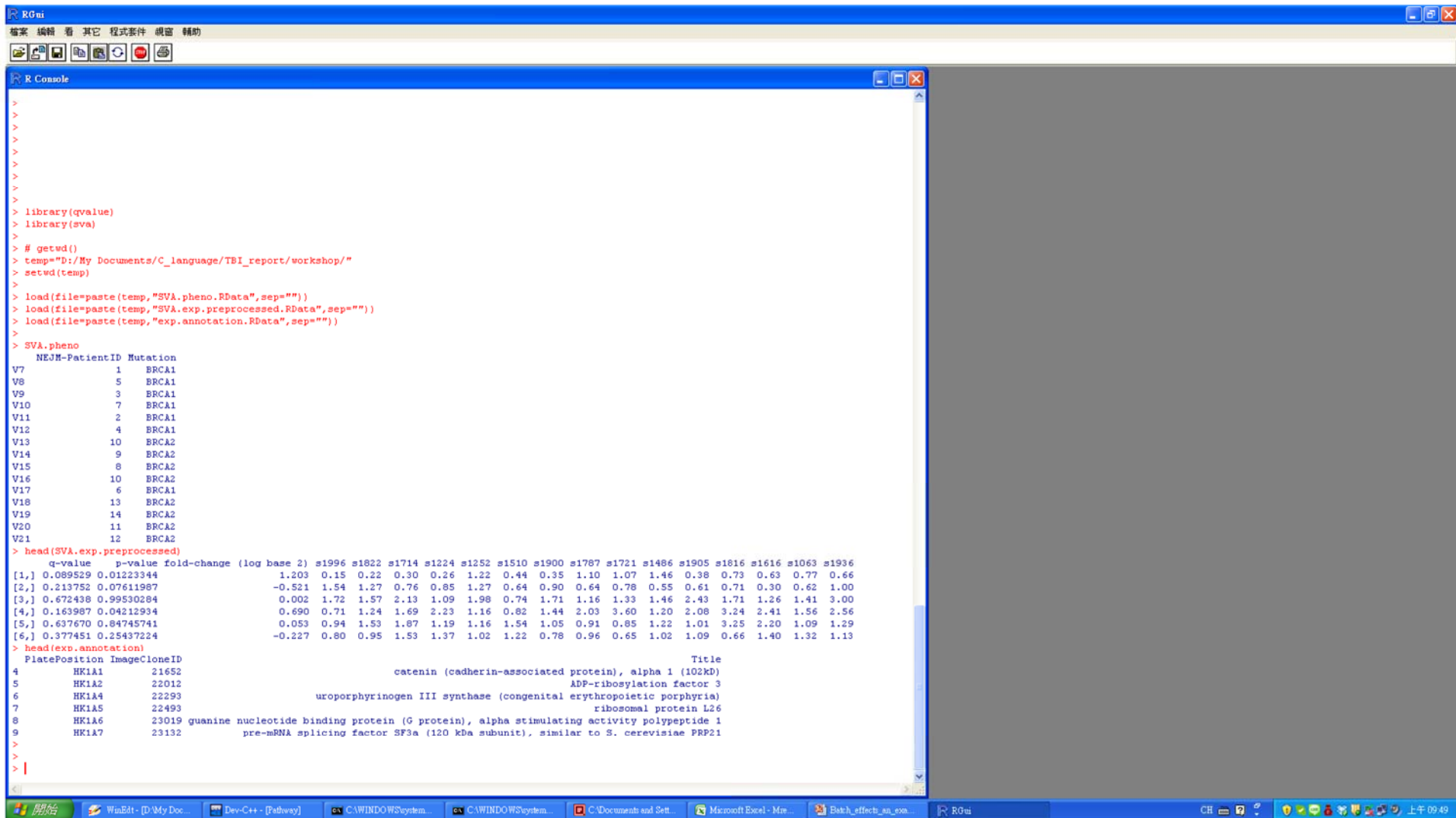
qsummary(qValuesObj.before)

modSv=cbind(mod,svaObj$sv[,1])
mod0Sv=cbind(mod0,svaObj$sv[,1])

pValues.after=f.pvalue(log2(SVA.exp.preprocessed[,-1:-3]),modSv,mod0Sv)
qValuesObj.after<-qvalue(pValues.after)

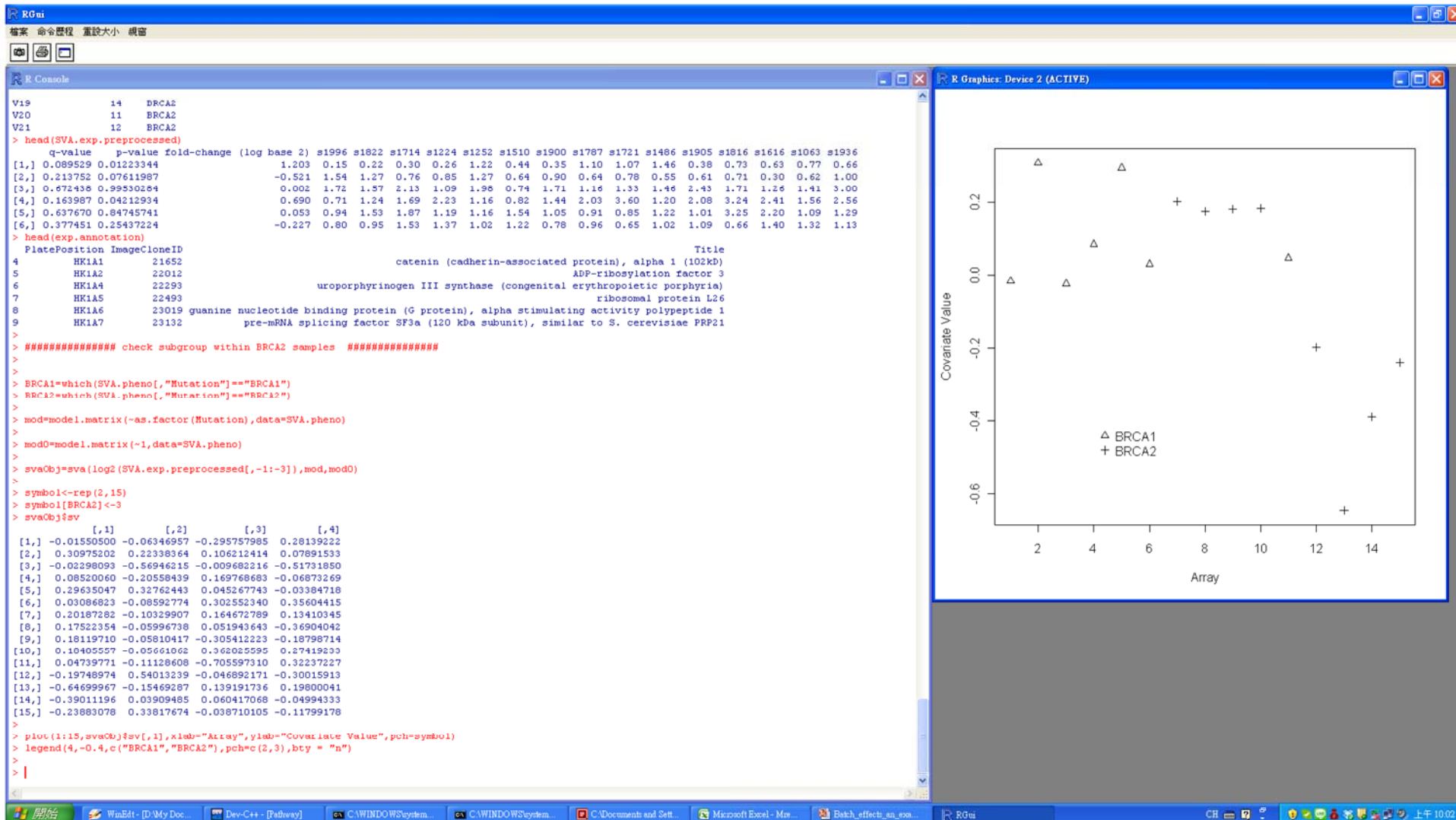
qsummary(qValuesObj.after)
```

Appendix: print screen (1)

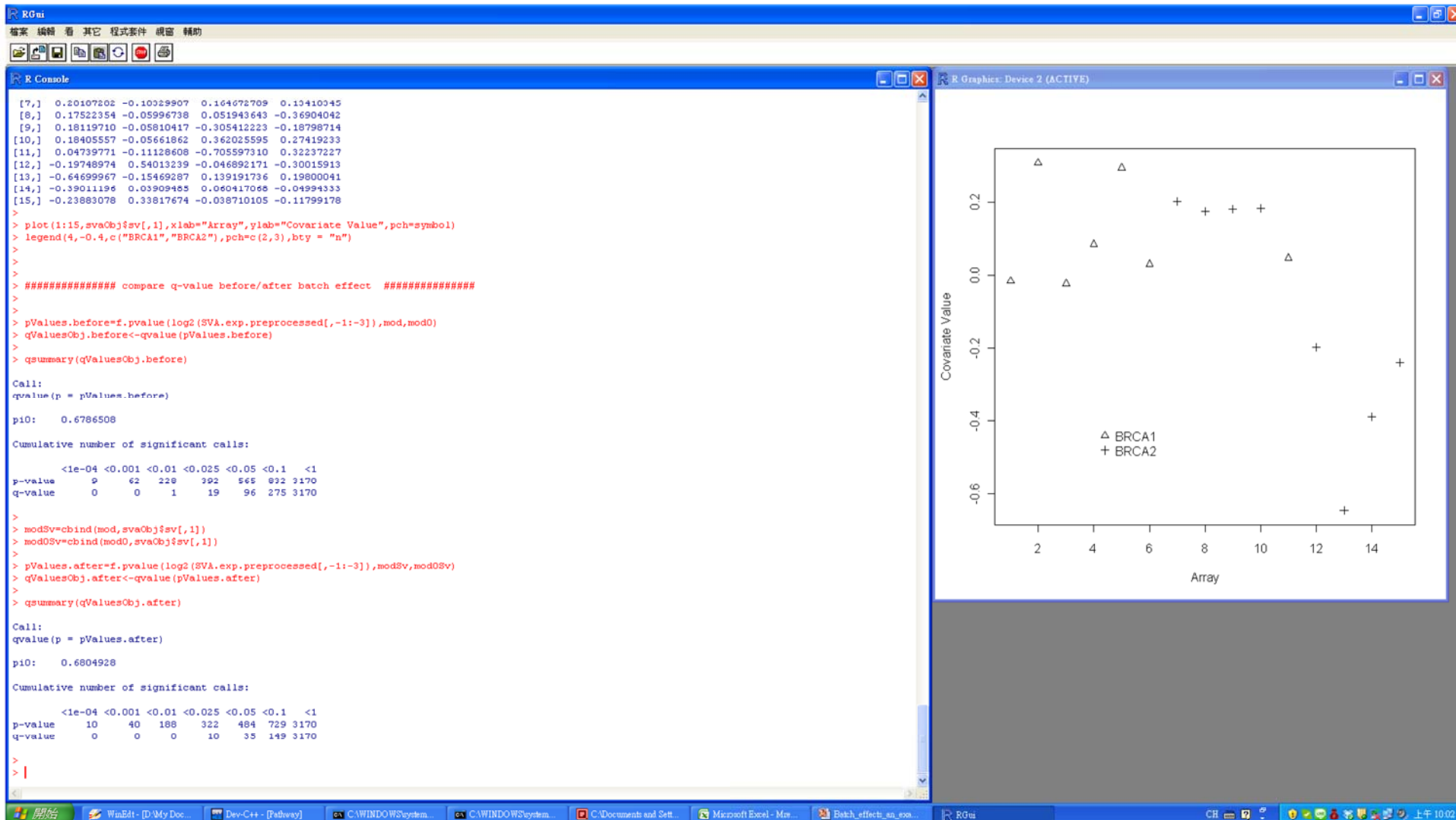


```
>
>
>
>
>
> library(qvalue)
> library(sva)
>
> # getwd()
> temp="D:/My Documents/C_language/TBI_report/workshop/"
> setwd(temp)
>
> load(file=paste(temp,"SVA.pheno.RData",sep=""))
> load(file=paste(temp,"SVA.exp.preprocessed.RData",sep=""))
> load(file=paste(temp,"exp.annotation.RData",sep=""))
>
> SVA.pheno
  NEJM-PatientID Mutation
V7              1  BRCA1
V8              5  BRCA1
V9              3  BRCA1
V10             7  BRCA1
V11             2  BRCA1
V12             4  BRCA1
V13            10  BRCA2
V14             9  BRCA2
V15             8  BRCA2
V16            10  BRCA2
V17             6  BRCA1
V18            13  BRCA2
V19            14  BRCA2
V20            11  BRCA2
V21            12  BRCA2
> head(SVA.exp.preprocessed)
  q-value p-value fold-change (log base 2) s1996 s1822 s1714 s1224 s1252 s1510 s1900 s1787 s1721 s1486 s1905 s1816 s1616 s1063 s1936
[1,] 0.089529 0.01223344      1.203 0.15 0.22 0.30 0.26 1.22 0.44 0.35 1.10 1.07 1.46 0.38 0.73 0.63 0.77 0.66
[2,] 0.213752 0.07611987     -0.521 1.54 1.27 0.76 0.85 1.27 0.64 0.90 0.64 0.78 0.55 0.61 0.71 0.30 0.62 1.00
[3,] 0.672438 0.99530284      0.002 1.72 1.57 2.13 1.09 1.98 0.74 1.71 1.16 1.33 1.46 2.43 1.71 1.26 1.41 3.00
[4,] 0.163987 0.04212934      0.690 0.71 1.24 1.69 2.23 1.16 0.82 1.44 2.03 3.60 1.20 2.08 3.24 2.41 1.56 2.56
[5,] 0.637670 0.84745741      0.053 0.94 1.53 1.87 1.19 1.16 1.54 1.05 0.91 0.85 1.22 1.01 3.25 2.20 1.09 1.29
[6,] 0.377451 0.25437224     -0.227 0.80 0.95 1.53 1.37 1.02 1.22 0.78 0.96 0.65 1.02 1.09 0.66 1.40 1.32 1.13
> head(exp.annotation)
  PlatePosition ImageCloneID Title
4             HK1A1      21652 catenin (cadherin-associated protein), alpha 1 (102kD)
5             HK1A2      22012 ADP-ribosylation factor 3
6             HK1A4      22293 uroporphyrinogen III synthase (congenital erythropoietic porphyria)
7             HK1A5      22493 ribosomal protein L26
8             HK1A6      23019 guanine nucleotide binding protein (G protein), alpha stimulating activity polypeptide 1
9             HK1A7      23132 pre-mRNA splicing factor SF3a (120 kDa subunit), similar to S. cerevisiae PRP21
>
>
>
```

Appendix: print screen (2)



Appendix: print screen (3)



Appendix: print screen (4)

