

GEMSiRV: A software platform for GEnome-scale Metabolic models Simulation, Reconstruction and Visualization

全基因規模的代謝網路建構、模擬與視覺化
平台之介紹與實際操作

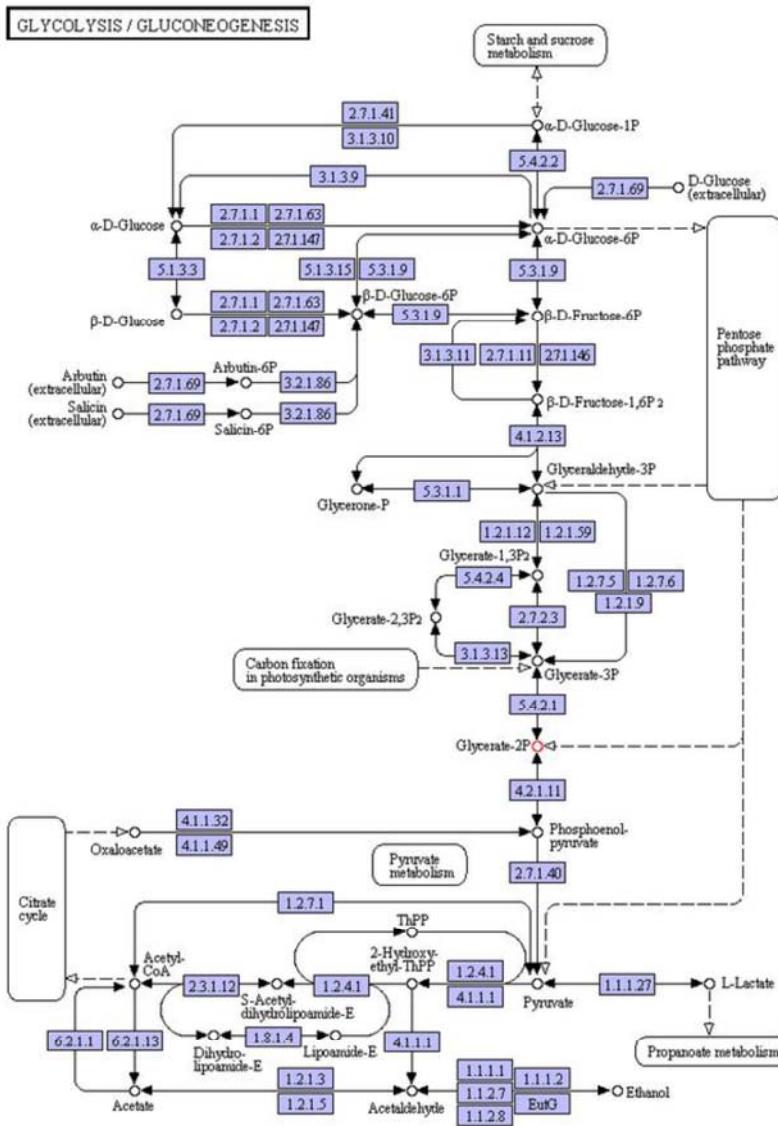
Yu-Chieh Liao, Ph.D.
Assistant Investigator
National Health Research Institutes

<http://sb.nhri.org.tw/GEMSiRV>

How to simulate a model?

WHAT IS A METABOLIC NETWORK?

KEGG pathway



00010 5/31/12
(c) Kanehisa Laboratories

- Enzyme catalyze metabolic reactions
- Reactants and products are involved in a reaction

E.g.



REACTION: R00658

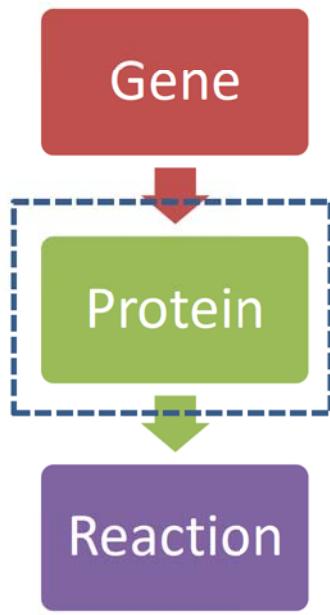
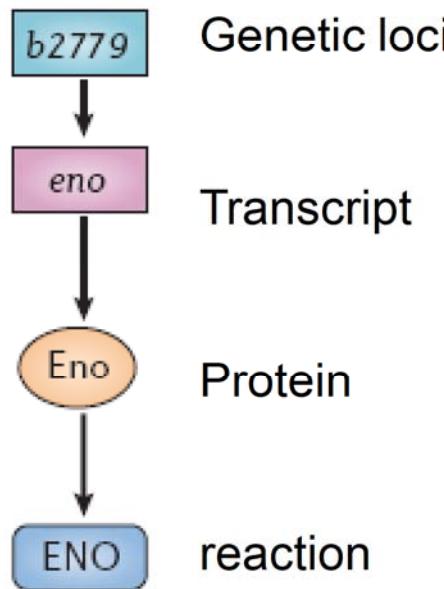
Entry	R00658	Reaction
Name	2-phospho-D-glycerate hydro-lyase (phosphoenolpyruvate-forming)	
Definition	2-Phospho-D-glycerate \leftrightarrow Phosphoenolpyruvate + H ₂ O	
Equation	C00631 \leftrightarrow C00074 + C00001	
RPair	RP01033_C00074_C00631 main RP06256_C00001_C00631 leave	
Enzyme	4.2.1.11	
Pathway	rn00010 Glycolysis / Gluconeogenesis rn00680 Methane metabolism rn01100 Metabolic pathways rn01110 Biosynthesis of secondary metabolites rn01120 Microbial metabolism in diverse environments rn01230 Biosynthesis of amino acids	
Orthology	KO1689 enolase [EC:4.2.1.11]	

http://www.genome.jp/kegg-bin/show_pathway?ko00010

What is in a metabolic network?

Abbreviation	Glycolytic reactions	Genes
HEX1	[c]GLC + ATP → G6P + ADP + H	glk
PGI	[c]G6P ↔ F6P	pgi
PFK	[c]ATP + F6P → ADP + FDP + H	pfkA, pfkB
FBA	[c]FDP ↔ DHAP + G3P	fbaA, fbaB
TPI	[c]DHAP ↔ G3P	tpiA
GAPD	[c]G3P + NAD + Pi ↔ 13DPG + H + NADH	gapA, gapC1, gapC2
PGK	[c]13DPG + ADP ↔ 3PG + ATP	pgk
PGM	[c]3PG ↔ 2PG	qpmA, qpmB
ENO	[c]2PG ↔ H ₂ O + PEP	eno
PYK	[c]ADP + H + PEP → ATP + PYR	pykA, pykF

Reactant → Product
2PG → H₂O and **PEP**
-1 → 1 1



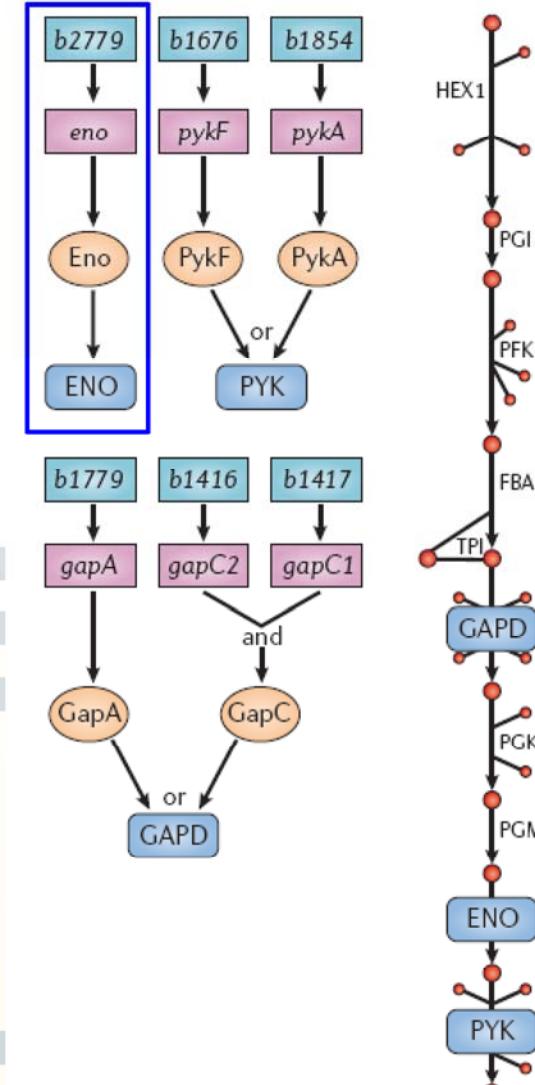
Box 3 | Assembly and representation

A list of charge and elementally balanced metabolic reactions can be represented in a stoichiometric matrix (S), where rows and columns correspond to metabolites and reactions and the elements are the stoichiometric coefficients. In genome-scale metabolic networks these stoichiometric matrices contain few non-zero elements, as relatively few metabolites participate in a given reaction. Connections between genes and reactions can be represented as gene–protein–reaction (GPR) associations by using Boolean rules or visualized using graphic images. In the GPR scheme, the first level (teal) corresponds to genetic loci, the second level (pink) to transcripts, the third level (orange) to functional proteins, and the fourth level (blue) to reactions. [c], cytoplasmic reactions.

Abbreviation	Glycolytic reactions	Genes
HEX1	[c]GLC + ATP \rightarrow G6P + ADP + H	glk
PGI	[c]G6P \leftrightarrow F6P	pgi
PFK	[c]ATP + F6P \rightarrow ADP + FDP + H	pfkA, pfkB
FBA	[c]FDP \leftrightarrow DHAP + G3P	fbaA, fbaB
TPI	[c]DHAP \leftrightarrow G3P	tpiA
GAPD	[c]G3P + NAD + PI \leftrightarrow 13DPG + H + NADH	gapA, gapC1, gapC2
PGK	[c]13DPG + ADP \leftrightarrow 3PG + ATP	pgk
PGM	[c]3PG \leftrightarrow 2PG	gapA, gapB
ENO	[c]2PG \leftrightarrow H ₂ O + PEP	eno
PYK	[c]ADP + H + PEP \rightarrow ATP + PYR	pykA, pykF

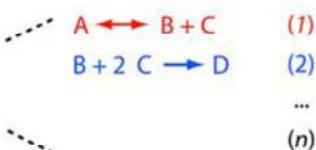
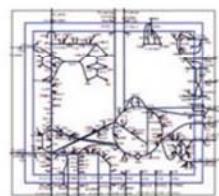
ATP	-1	0	-1	0	0	0	1	0	0	1
GLC	-1	0	0	0	0	0	0	0	0	0
ADP	1	0	1	0	0	0	-1	0	0	-1
G6P	1	-1	0	0	0	0	0	0	0	0
H	1	0	1	0	0	1	0	0	0	-1
F6P	0	1	-1	0	0	0	0	0	0	0
FDP	0	0	1	-1	0	0	0	0	0	0
DHAP	0	0	0	1	-1	0	0	0	0	0
G3P	0	0	0	1	1	-1	0	0	0	0
NAD	0	0	0	0	0	-1	0	0	0	0
PI	0	0	0	0	0	-1	0	0	0	0
13DPG	0	0	0	0	0	1	-1	0	0	0
NADH	0	0	0	0	0	1	0	0	0	0
3PG	0	0	0	0	0	0	1	-1	0	0
2PG	0	0	0	0	0	0	0	1	-1	0
PEP	0	0	0	0	0	0	0	0	1	-1
H ₂ O	0	0	0	0	0	0	0	0	1	0
PYR	0	0	0	0	0	0	0	0	0	1

HEX1	PGI	PFK	FBA	TPI	GAPD	PGK	PGM	ENO	PYK
------	-----	-----	-----	-----	------	-----	-----	-----	-----



Flux balance analysis

a Curate metabolic reactions



b Formulate S matrix

	Reactions					
Metabolites	A	B	C	D	...	n
A	1	2	...			
B	-1	1	-1			
C	1	-2				
D		1				
...						
m						

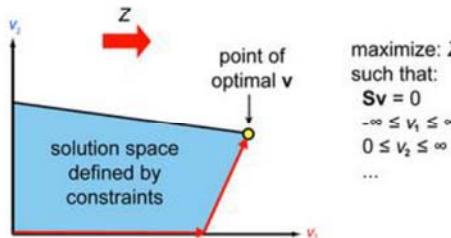
c Apply mass balance constraints

$$\begin{matrix} S (m \times n) & v (n \times 1) \\ \begin{bmatrix} -1 \\ 1 \\ -1 \\ 1 \end{bmatrix} & \begin{bmatrix} v_1 \\ v_2 \\ \dots \\ v_n \end{bmatrix} \end{matrix} * = 0 \rightarrow \begin{matrix} m \text{ mass balance equations} \\ -v_1 + \dots = 0 \\ v_1 - v_2 + \dots = 0 \\ v_1 - 2v_2 + \dots = 0 \\ v_2 + \dots = 0 \\ \dots \end{matrix}$$

d Define objective function Z

$$Z = \begin{matrix} v (n \times 1) \\ \begin{bmatrix} v_1 \\ v_2 \\ \dots \\ v_n \end{bmatrix} \end{matrix} * \begin{matrix} c^T (1 \times n) \\ \begin{bmatrix} 1 & 0 & \dots & 0 \end{bmatrix} \end{matrix} \rightarrow \begin{matrix} \text{sets reaction 1 as} \\ \text{the objective} \end{matrix}$$

e Optimize Z using linear programming



• Linear programming

Linear programs are problems that can be expressed in canonical

$$\text{maximize } c^T x$$

$$\text{subject to } Ax \leq b$$

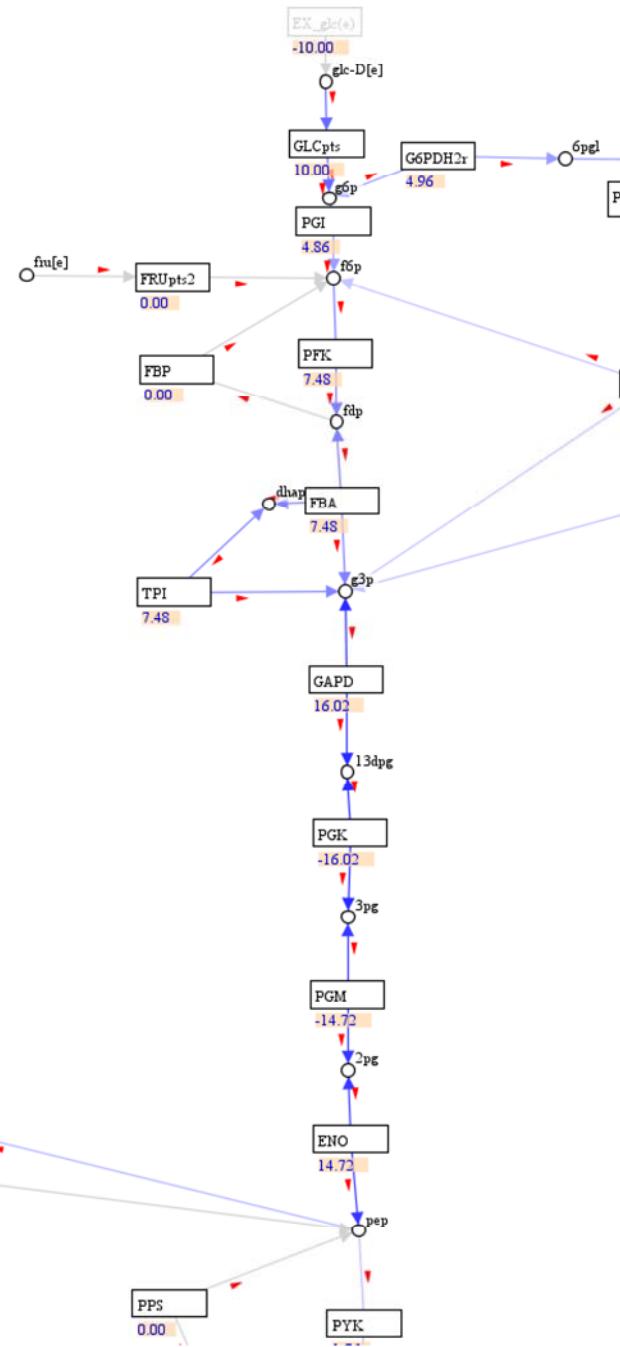
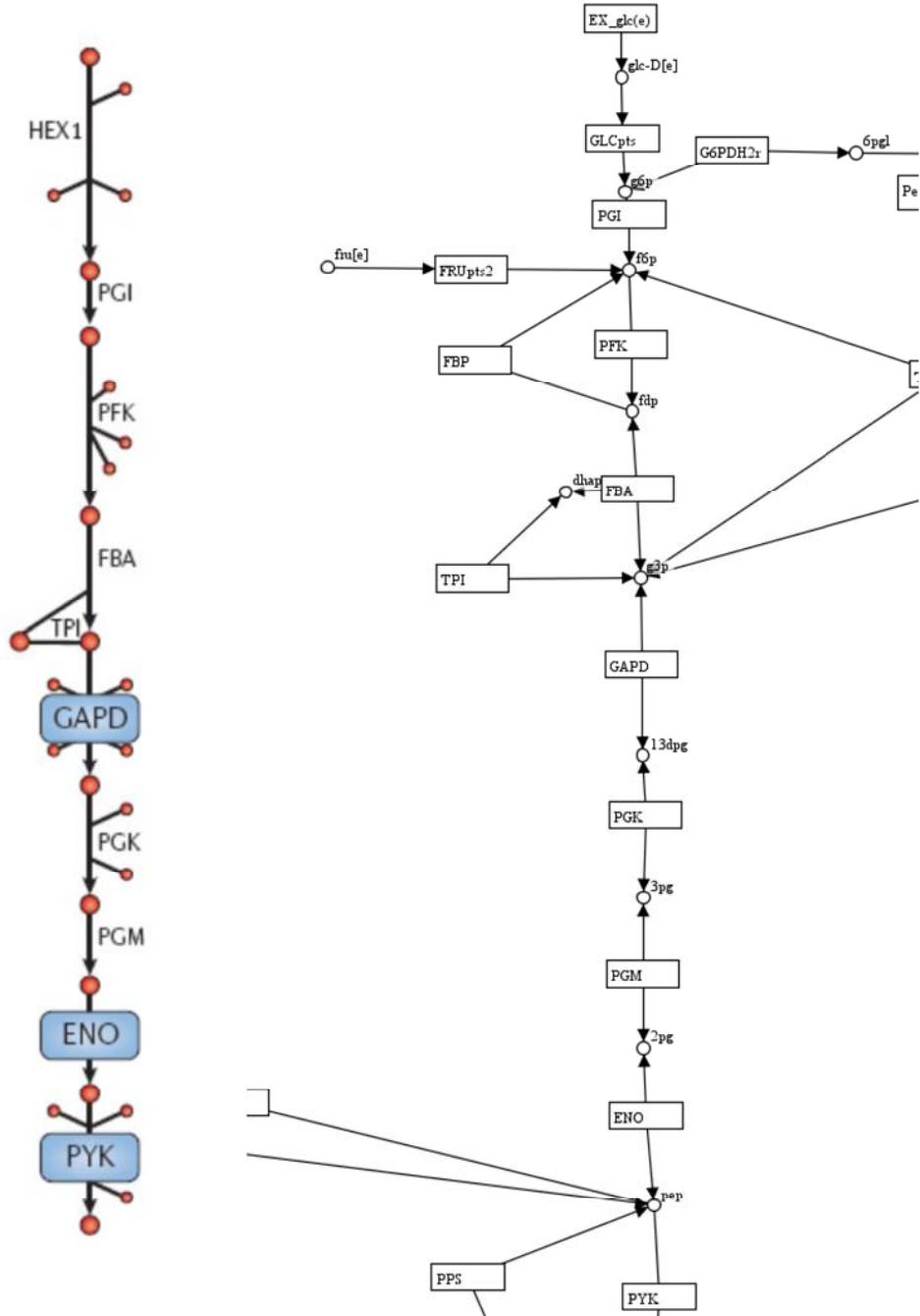
$$\text{and } x \geq 0$$

Biomass:

(1.496) 3pg + (3.7478) accoa + (59.81) atp +
(0.361) e4p + (0.0709) f6p + (0.129) g3p +
(0.205) g6p + (0.2557) gln-L + (4.9414) glu-L
+ (59.81) h2o + (3.547) nad + (13.0279)
nadph + (1.7867) oaa + (0.5191) pep +
(2.8328) pyr + (0.8977) r5p

-->

(59.81) adp + (4.1182) akg + (3.7478) coa +
(59.81) h + (3.547) nadh + (13.0279) nadp +
(59.81) pi



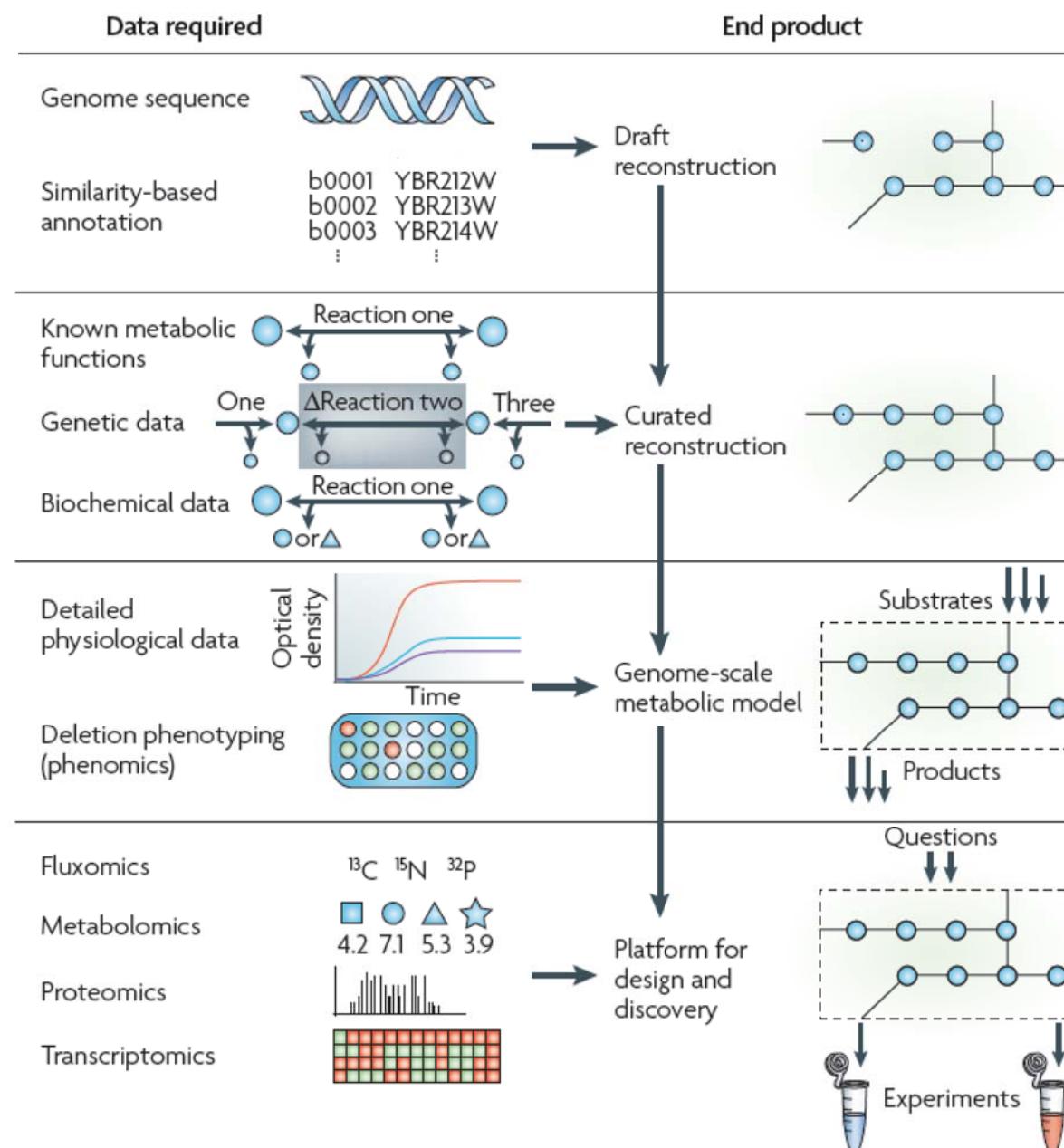


Figure 1 | Phases and data used to generate a metabolic reconstruction.

Systems biology

Advance Access publication May 4, 2012

GEMSiRV: a software platform for GEnome-scale metabolic model simulation, reconstruction and visualization

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Associate Editor: Trey Ideker

MrBac (<http://sb.nhri.org.tw/MrBac>)

A web server to draft

Metabolic network reconstructions for Bacteria
Mr Bac



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*Select a query and a reference species: [[Example](#)]

Query Species:	Reference Species:
Acaryochloris marina MBIC11017	Acaryochloris marina MBIC11017
Acholeplasma laidlawii PG-8A	Acholeplasma laidlawii PG-8A
Acidiphilium cryptum JF-5	Acidiphilium cryptum JF-5
Acidithiobacillus ferrooxidans ATCC 23270	Acidithiobacillus ferrooxidans ATCC 23270
Acidithiobacillus ferrooxidans ATCC 53993	Acidithiobacillus ferrooxidans ATCC 53993
Acidobacteria bacterium Ellin345	Acidobacteria bacterium Ellin345
Acidobacterium capsulatum ATCC 51196	Acidobacterium capsulatum ATCC 51196
Acidothermus cellulolyticus 11B	Acidothermus cellulolyticus 11B
Acidovorax citrulli AAC00-1	Acidovorax citrulli AAC00-1
Acidovorax sp. JS42	Acidovorax sp. JS42

*Choose Blast Settings: [[Default](#) / [Select All](#)]

E value: & Percent Identity: %

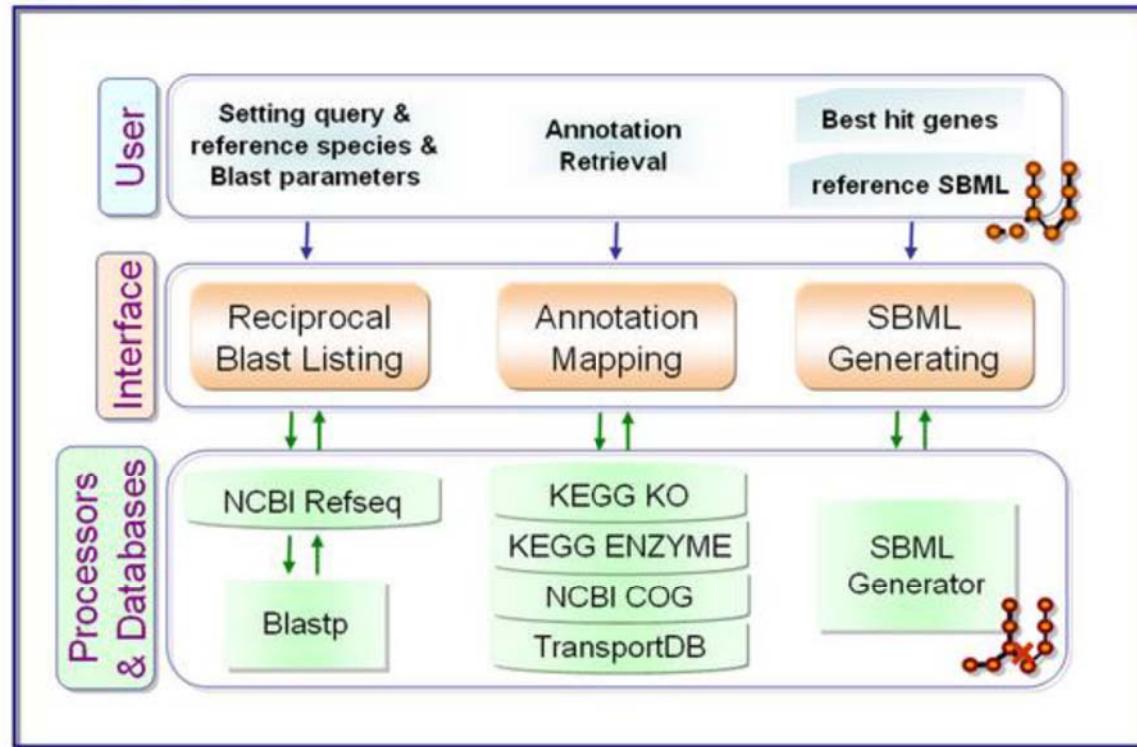
Choose the blast columns to be shown: [?](#)

Ref Query Perclden MLen mismatches gap openings q.start q.end r.start r.end EVal BitScore

* indicates required input.

Submit

Flow Chart



Metabolic Network Draft Generation

Draft metabolic network generation requires a reference SBML file and the list of reciprocal blast best hits as inputs to proceed.

By taking these two inputs, we are assuming that query ORFs in the best hit list have the same functions as their corresponding reference ORFs. The generator only keeps reactions, in which all associated reference ORFs under "AND" condition and partial reference ORFs under "OR" condition have best hit matches to query ORFs. Only when those two criteria are met, the reference ORF id are replaced by corresponding query id, and the metabolites involved in those reactions are kept in the draft SBML file.

The draft in SBML format can be saved as the final output after query is submitted and can be further curated and edited manually in flux balance analysis softwares such as [COBRA](#).

Upload a reference SBML file ([sample_iAF1260](#)) [?](#)

SBML File:

Upload a one to one gene pair file ([sample_SLT2vsECO](#)) [?](#)

Gene Pair File:

Type in a model name

New Model Name:

```
<?xml version="1.0" encoding="UTF-8"?>
<sbml level="2" version="1" >
  <model id="Ec_iAF1260" name="Ec_iAF1260" >
    <listOfUnitDefinitions >
      <unitDefinition id="mmol_per_gDW_per_hr" >
        <listOfUnits >
          <unit kind="mole" scale="-3" />
          <unit exponent="-1" kind="gram" />
          <unit exponent="-1" kind="second" multiplier=".00027777" />
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      </unitDefinition>
    </listOfUnitDefinitions>
    <listOfCompartments >
      <compartment id="Extra_organism" />
      <compartment id="Periplasm" outside="Extra_organism" />
      <compartment id="Cytosol" outside="Periplasm" />
    </listOfCompartments>
    <listOfSpecies >
      <species boundaryCondition="false" charge="-2" compartment="Cytosol" id="M_10fthf_c" name="M_10_Formyltetrahydrofolate_C2">
      <species boundaryCondition="false" charge="0" compartment="Cytosol" id="M_12ppd_R_c" name="M_R_Propane_1_2_diol_C3H8O2">
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      <species boundaryCondition="false" charge="-4" compartment="Cytosol" id="M_13dpq_c" name="M_3_Phospho_D_glyceroyl_phospho">
      <species boundaryCondition="false" charge="0" compartment="Cytosol" id="M_14glucan_c" name="M_1_4_alpha_D_glucan_C36H62O3">
      <species boundaryCondition="false" charge="0" compartment="Periplasm" id="M_14glucan_p" name="M_1_4_alpha_D_glucan_C36H62O3">
      <species boundaryCondition="false" charge="2" compartment="Cytosol" id="M_15dap_c" name="M_1_5_Diaminopentane_C5H16N2" />
      <species boundaryCondition="false" charge="2" compartment="Periplasm" id="M_15dap_p" name="M_1_5_Diaminopentane_C5H16N2" />
      <species boundaryCondition="false" charge="0" compartment="Periplasm" id="M_lagpe120_p" name="M_1_Acyl_sn_glycero_3_phosp">
      <species boundaryCondition="false" charge="0" compartment="Periplasm" id="M_lagpe140_p" name="M_1_Acyl_sn_glycero_3_phosp">
      <species boundaryCondition="false" charge="0" compartment="Periplasm" id="M_lagpe141_p" name="M_1_Acyl_sn_glycero_3_phosp">
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      <species boundaryCondition="false" charge="0" compartment="Periplasm" id="M_lagpe181_p" name="M_1_Acyl_sn_glycero_3_phosp">
      <species boundaryCondition="false" charge="-1" compartment="Periplasm" id="M_lagpg120_p" name="M_1_Acyl_sn_glycero_3_phos">
      <species boundaryCondition="false" charge="-1" compartment="Periplasm" id="M_lagpg140_p" name="M_1_Acyl_sn_glycero_3_phos">
    </listOfSpecies>
  </model>
</sbml>
```

Starting materials for manual curation

Protein ID	Protein Name	Gene ID	Description	EC Number	Enzyme Class	Reaction	Substrate	Product	KEGG ID	Pathway	GO ID	GO Term	UniProt ID
STM1097..	hypothetical protein	1184453	1184863				136 K07127	-- Unclassified	-- -- --	R	Poorly General function		b1970
STM1098hpaC	4-hydroxyphenylacetate catabolism	1185708	1185196				170 K00484	4-hydroxy Metabolism	1.14.13.3 4-hydroxy Oxidoreductase	R	Poorly General function		
STM1099hpaB	4-hydroxyphenylacetate catabolism	1187288	1185726				520 K00483	4-hydroxy Metabolism	1.14.13.3 4-hydroxy Oxidoreductase	Q	METABC Secondary		
STM1100hpaR	4-hydroxyphenylacetate catabolism	1187946	1187506				146 ..	-- -- --	-- -- --	K	INFORM, Transcript		
STM1101hpaG	4-hydroxyphenylacetate catabolism	1188221	1189510				429 K05921	5-oxopent Metabolism	4.1.1.68;; 5-oxopent Lyases; C:5-oxopent Q		METABC Secondary		
STM1102hpaE	4-hydroxyphenylacetate catabolism	1189507	1190973				488 K00151	5-carboxy Metabolism	1.2.1.60 5-carboxy Oxidoreductase	C	METABC Energy pr		
STM1103hpaD	4-hydroxyphenylacetate catabolism	1190975	1191826				283 K00455	3,4-dihyd Metabolism	1.13.11.1;; 3,4-dihyd Oxidoreductase	S	Poorly Function		
STM1104hpaF	4-hydroxyphenylacetate catabolism	1191836	1192216				126 K01826	5-carboxy Metabolism	5.3.3.10 5-carboxy Isomerase	E	METABC Amino aci		
STM1105hpaH	4-hydroxyphenylacetate catabolism	1192359	1193162				267 K02509	2-oxo-hep Metabolism	4.2.1. - Lyases; C	Q	METABC Secondary		
STM1106hpaI	4-hydroxyphenylacetate catabolism	1193173	1193964				263 K02510	2,4-dihyd Metabolism	4.1.2. - Lyases; C	G	METABC Carbohydr		
STM1107hpaX	4-hydroxyphenylacetate catabolism	1194036	1195412				458 K02511	MFS trans Protein Fa	-- -- --	G; E; P	METABC Carbohydr Secondary	The Major 4-hydroxy	
STM1108hpaA	4-hydroxyphenylacetate catabolism	1195422	1196318				298 K02508	AraC fam Protein Fa	-- -- --	K	INFORM, Transcript		
STM1109..	putative periplasmic protein	1196332	1197270				312 ..	-- -- --	-- -- --	R	Poorly General function		

- We can identify, for instance, the metabolic genes that are present in SLT2 but absent in ECO.
 - Examples include the cob (STM2016-STM2035), hpa (STM1098-STM1108) and pdu (STM2037-STM2058).
 - These genes have been reported to confer SLT2 the abilities to synthesize cobalamin and to utilize 4-hydroxylphenylacetate and propanediol.



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GEMSiRV

A software platform for GEnome-scale Metabolic Simulation, Reconstruction and Visualization.

This work has been published in Bioinformatics (Pubmed®). If you have any question, please do not hesitate to contact the author by emailing to sysbio@nhri.org.tw.

Start GEMSiRV®. An example model (*E. coli* textbook model) and its map can be directly imported by click on the [Help to Import an Example](#).

Genome-scale metabolic network models have become an indispensable part of the increasingly important field of systems biology.

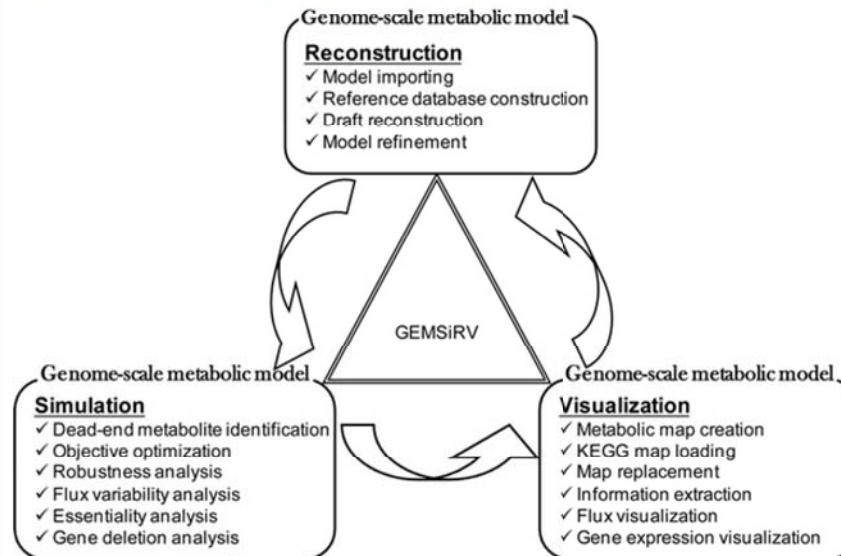
Metabolic systems biology studies usually include three major components – network model construction, objective- and experiment-guided model editing and visualization, and simulation studies based mainly on flux balance analyses.

Here we present a software platform, GEMSiRV, to provide functionalities of easy metabolic network drafting and editing, amenable network visualization for experimental data integration, and flux balance analysis tools for simulation studies.

GEMSiRV is an open-source software for building users' metabolic systems biology project, it provides interactive features in model management, simulation, visualization and integration of omics data.

Furthermore, all of the GEMSiRV-generated metabolic models and analysis results, including projects in progress, can be easily exchanged in the research community. GEMSiRV is a powerful integrative resource that may facilitate the development of systems biology studies.

A schematic overview of the GEMSiRV outlines the key features implemented in the GEMSiRV.



Acknowledgements

- Dr. Chao A. Hsiung
- Dr. Feng-Chi Chen
- Mr. Ming-Hsin Tsai
- Ms. Julie Chih-Yu Chen
- Ms. Yueh-Hsia Tang
- **National Health Research Institutes**
- **National Science Council**

Practices

Metabolic models

- Gene-protein-reaction association
 - Excel model
- Gene/protein information is not necessary
 - Excel or xml model

<http://sb.nhri.org.tw/GEMSiRV/en/Manual>

▪ Dead-end metabolite identification (Step-by-step

Select a metabolic model and a map (if you have). GEMSiRV can identify dead-end metabolites and tag them with crosses in the map.

▪ Objective optimization (Step-by-step

Select a metabolic model and a map (if you have) for objective optimization, the flux result can be visualized in the map.

▪ Flux variability analysis (Step-by-step

Select a metabolic model and a map (if you have) for flux variability analysis, the min and max fluxes of reaction can be plotted in the map and the blocked reaction are tagged with crosses.

▪ Robustness analysis (Step-by-step

Select the reactions of interest in a model to see how sensitive the objective is to the particular reactions.

▪ Essentiality analysis (Step-by-step

Select a metabolic model for essentiality analysis, the computational essential genes or reactions can be identified.

▪ Gene deletion analysis (Step-by-step

Select a metabolic model for gene deletion analysis, the gene-deletion model can be saved and imported for further evaluation.

