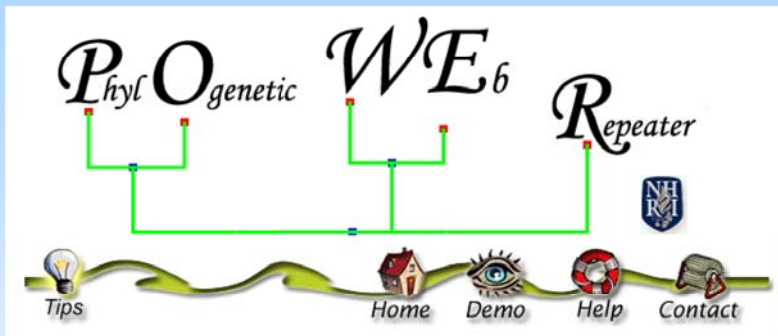


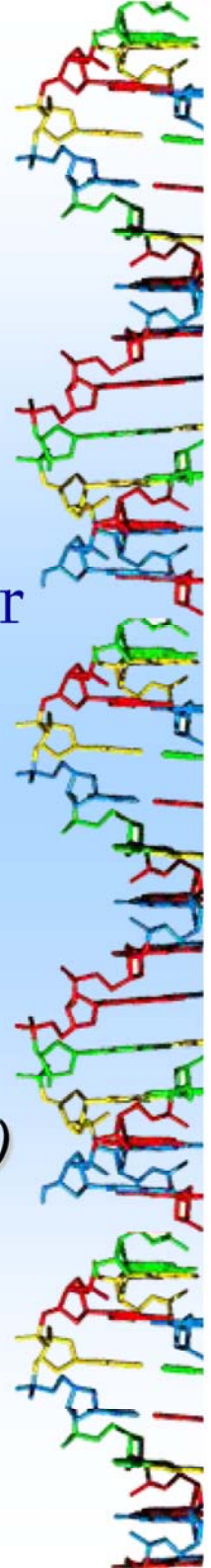
POWER: PhylOgenetic WEb Repeater

- An integrated and user-optimized framework for biomolecular phylogenetic analysis



Speaker: Chieh -Hua Lin (林介華)

mammer@nhri.org.tw



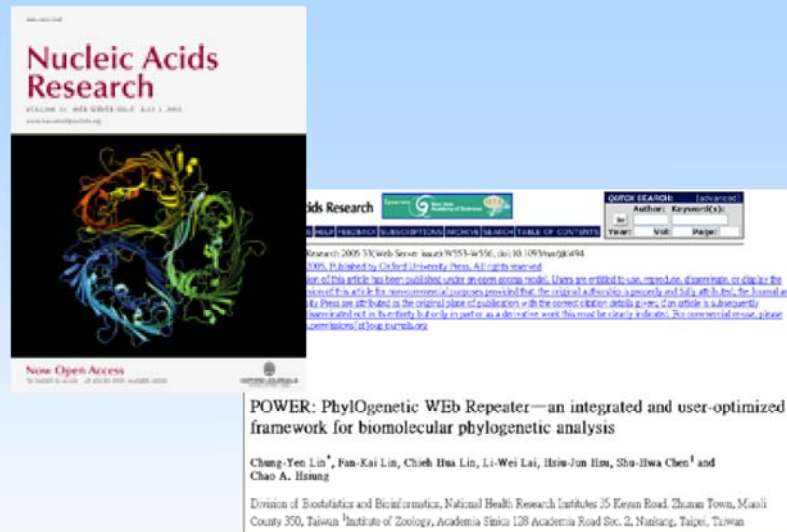
Phylogenetic Analysis



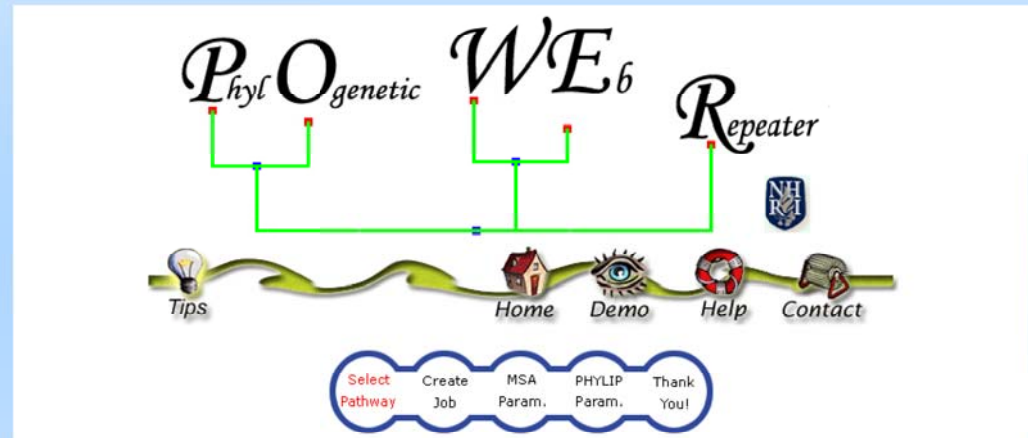
- Based on observable evidence to **estimate** or **infer** their evolution relationship
- **Phylogenetic tree** can be used to represent the historical relationships of groups of organisms, not similarities
 - a graphical representation of the evolution relations among organisms or taxa. The taxa can be species, populations, individuals or genes.

POWER: Phylogenetic Web Repeater - An integrated and user-optimized framework for biomolecular phylogenetic analysis

<http://power.nhri.org.tw>



Nucl. Acids Res. 2005 33: W553-W556



The Phylogenetic Web Repeater (POWER) allows users performing phylogenetic analysis with molecular data by most programs of PHYLIP package repeatedly. POWER provide two pipelines to process the analysis. One of them includes multiple sequence alignment (MSA) at the beginning of the pipeline whereas the other begin phylogenetic analysis with aligned sequence.

Please start your analysis by selecting the pipeline and the data type:

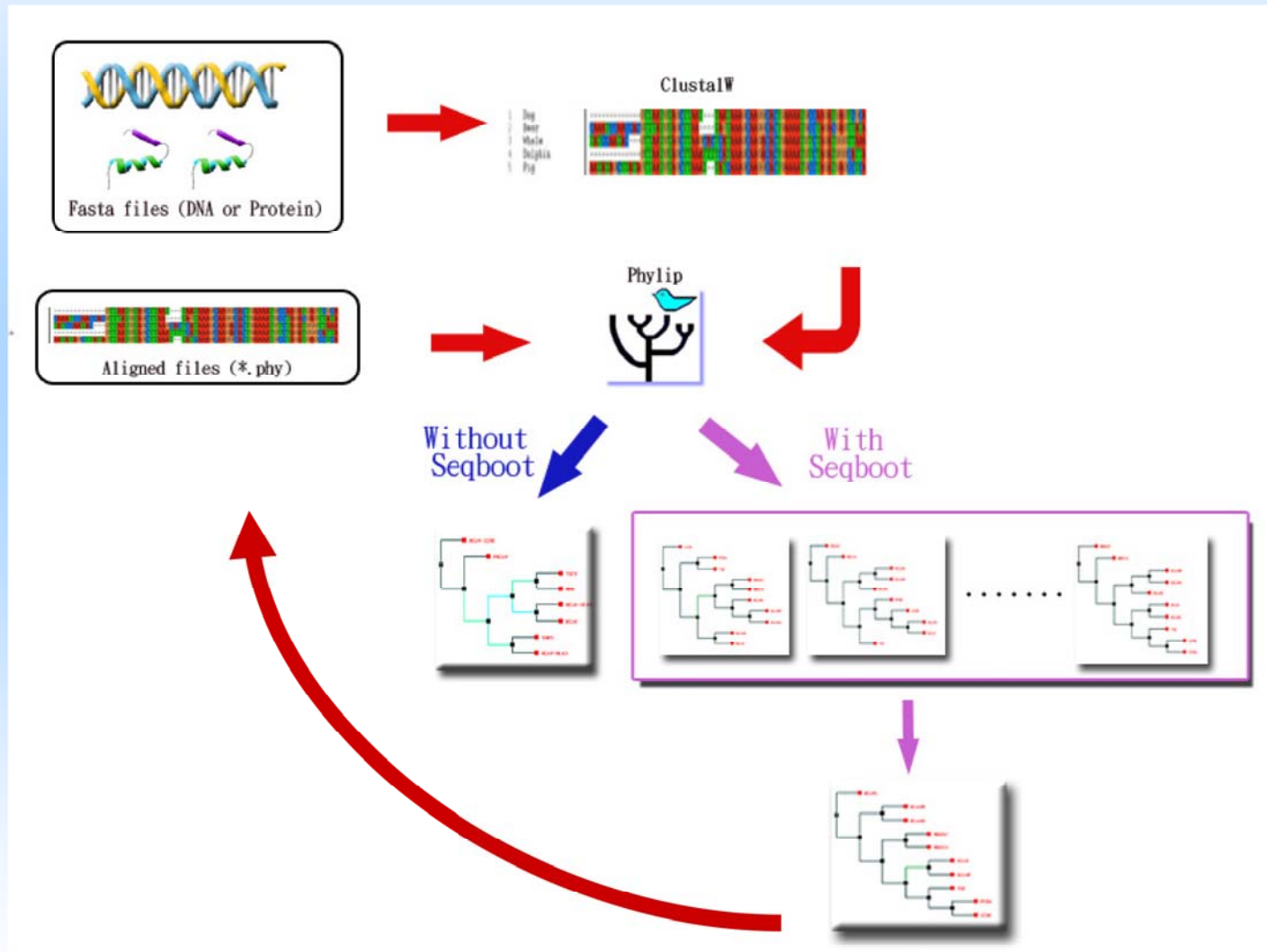
Pipeline	<input type="radio"/> MSA + Phylogenetic Analysis(Input the FASTA format)
	<input type="radio"/> Phylogenetic Analysis Only(Input the PHYLIP format)
Sequence Type	<input type="radio"/> DNA
	<input type="radio"/> Protein

Phylogenetic Analysis Steps

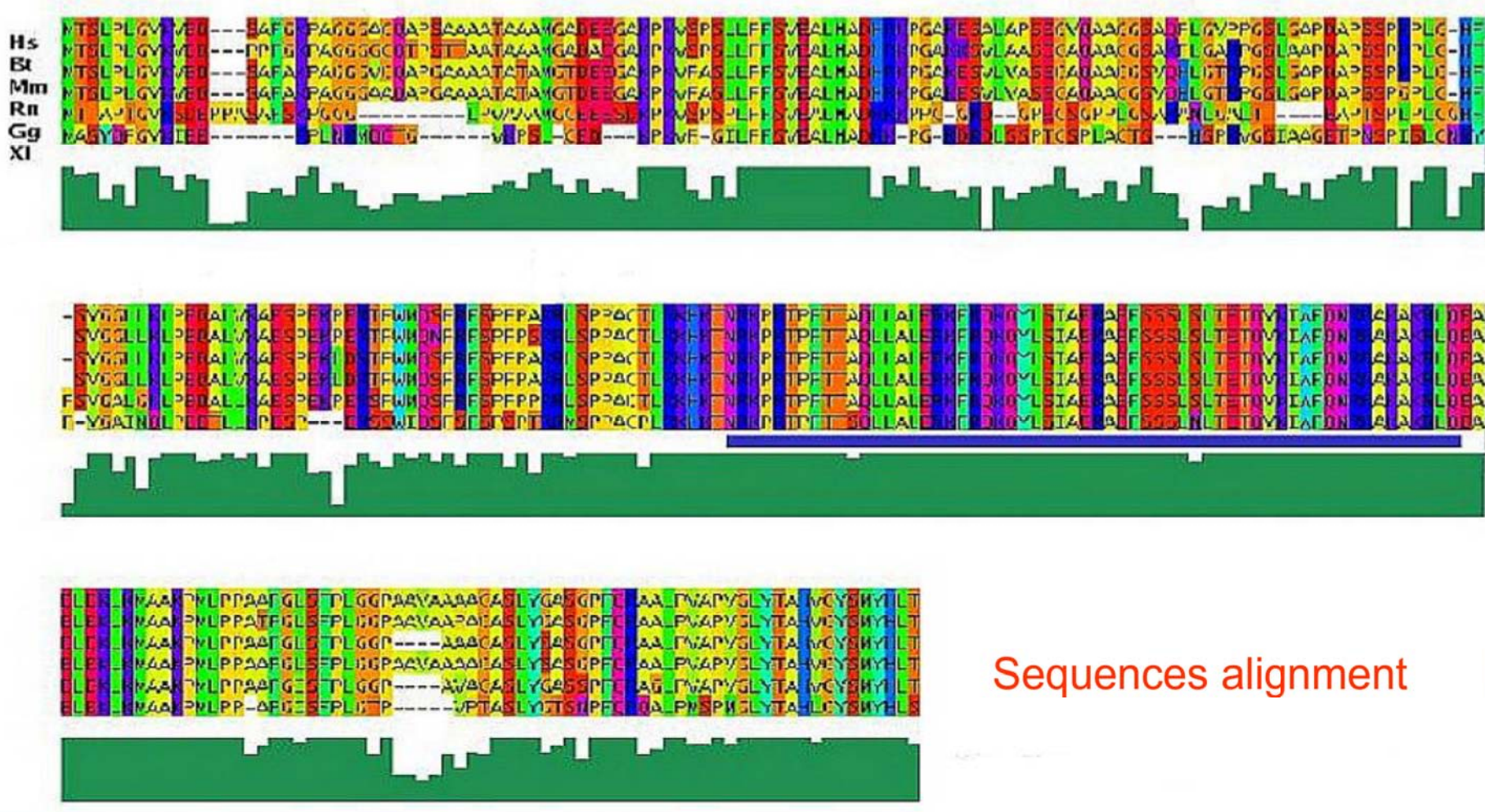


- Sequences alignment
- Build substitution model
- Construct phylogenetic tree
- Evaluate phylogenetic tree

PhylOgenetic Web Repeater (POWER)


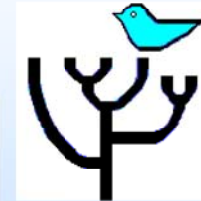








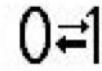
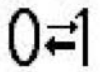
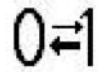




























Schema of POWER




Sequences alignment

Phylip



 dnainvar.exe	 dnaml.exe	 dnamlk.exe	 dnamove.exe	 dnapars.exe	 dnapenny.exe	 dollop.exe
 dolmove.exe	 dolpenny.exe	 drawgram.exe	 drawtree.exe	 factor.exe	 fitch.exe	 gendist.exe
 kitsch.exe	 mix.exe	 move.exe	 neighbor.exe	 pars.exe	 penny.exe	 proml.exe
 promlk.exe	 protdist.exe	 protpars.exe	 restdist.exe	 restml.exe	 retree.exe	 seqboot.exe
 treedist.exe	 font1	 font2	 font3	 font4	 font5	 font6
 neighbor	 protdis					

Interactive Interface for Phylip



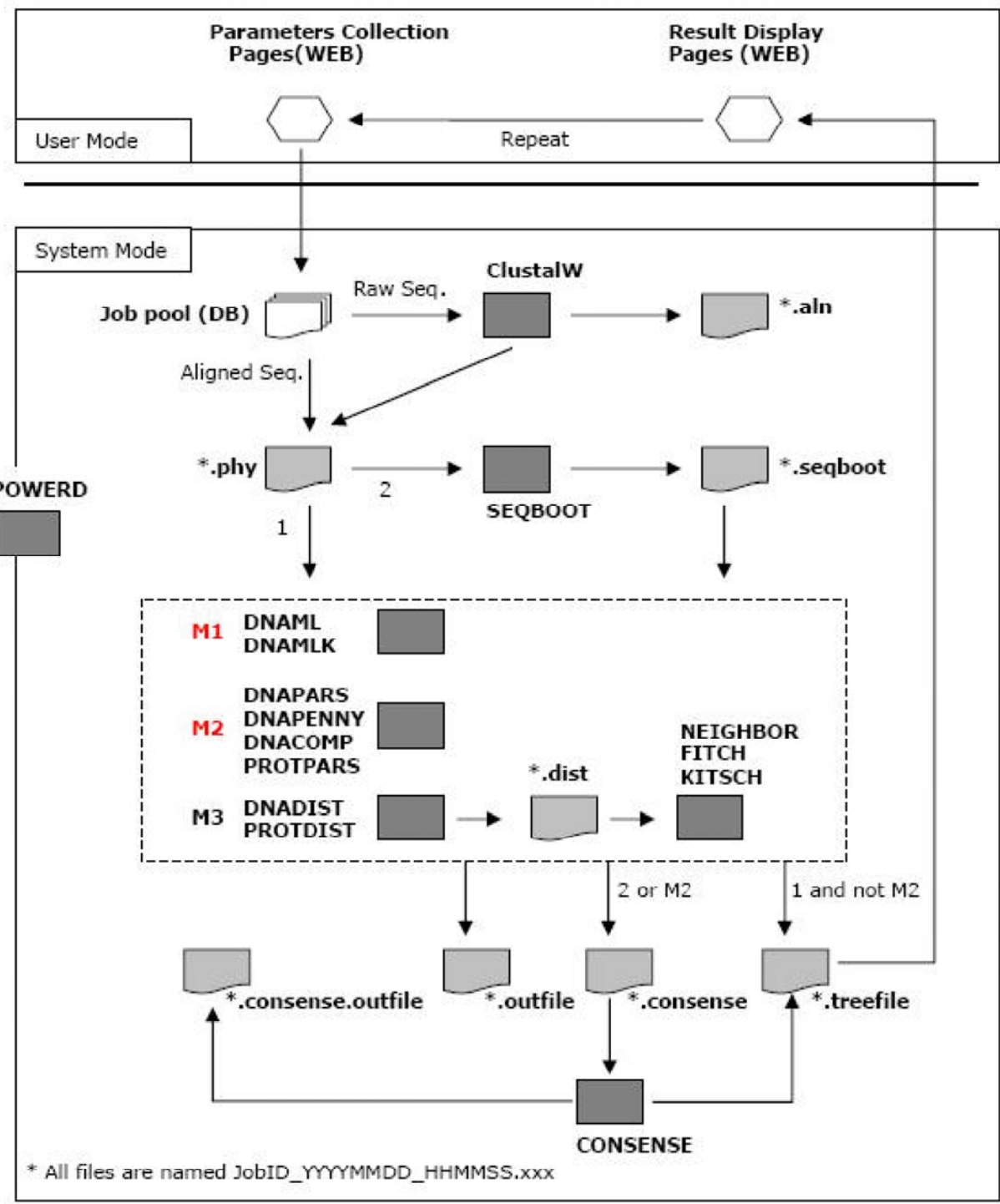
```
Nucleic acid sequence Maximum Likelihood method, version 3.6

Settings for this run:
U          Search for best tree?  Yes
T          Transition/transversion ratio:  2.0000
F          Use empirical base frequencies?  Yes
C          One category of sites?  Yes
R          Rate variation among sites?  constant rate
W          Sites weighted?  No
S          Speedier but rougher analysis?  Yes
G          Global rearrangements?  No
J          Randomize input order of sequences?  No. Use input order
O          Outgroup root?  No, use as outgroup species  1
M          Analyze multiple data sets?  No
I          Input sequences interleaved?  Yes
O          Terminal type (IBM PC, ANSI, none)?  ANSI
1          Print out the data at start of run  No
2          Print indications of progress of run  Yes
3          Print out tree  Yes
4          Write out trees onto tree file?  Yes
5          Reconstruct hypothetical sequences?  No

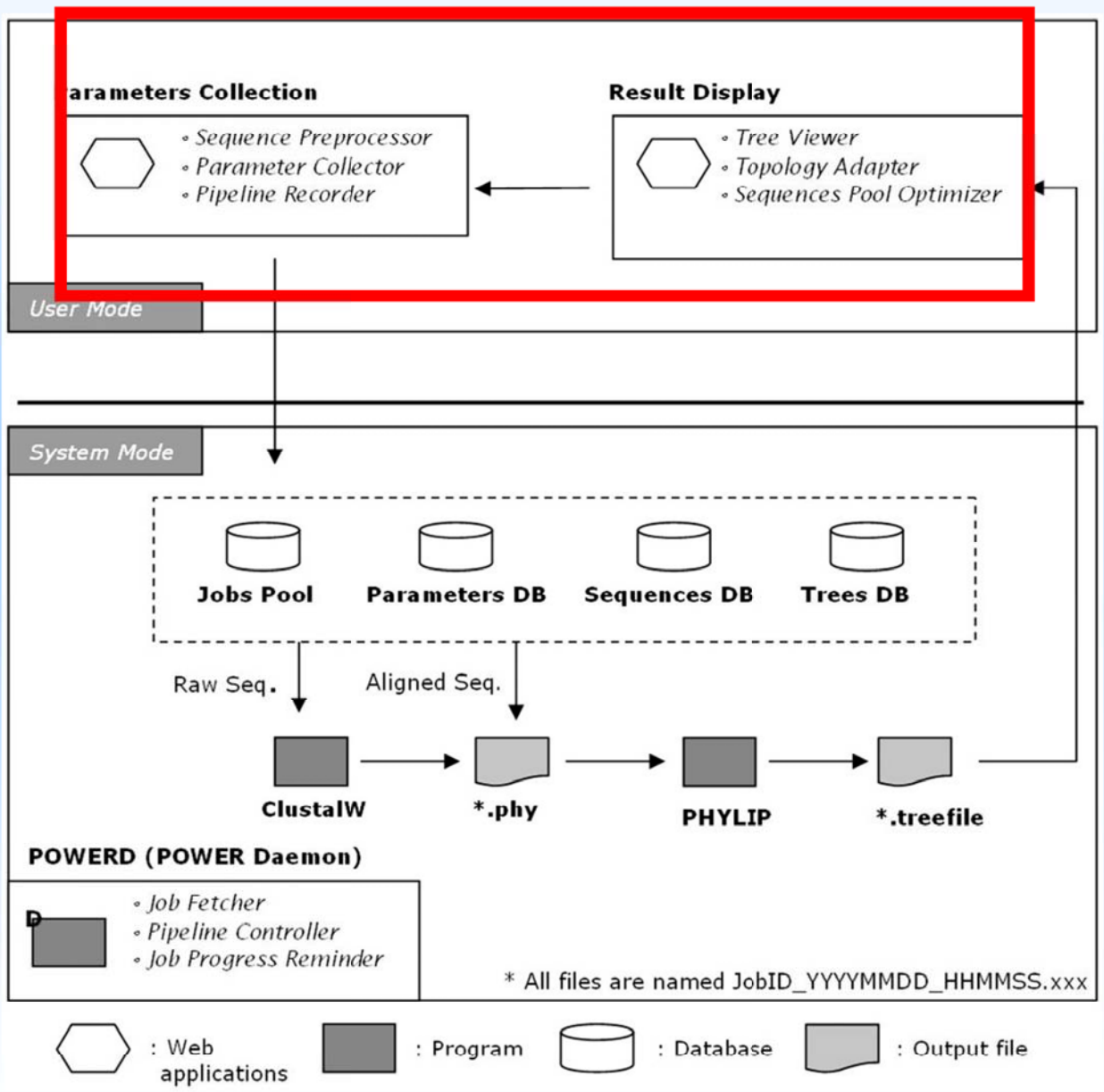
Y to accept these or type the letter for one to change
```

At this stage they do not have a mouse-windows interface for PHYLIP





 : web site
  : program
  : database
  : output file



Phylogenetic Analysis Step 1



- Sequences alignment
 - ClustalW, Jalview



Input file

- FASTA format: start with a symbol “>”
- The names of each sequence are not more than 10 characters.

❑ 1: [AF029248](#). Reports Mouse hepatitis v...[gi:2641127]

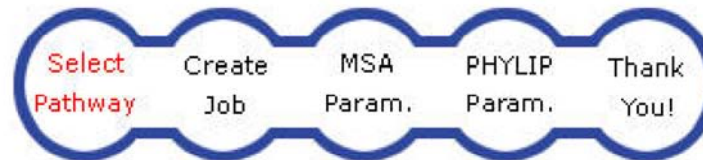
```
>gi|2641127|gb|AF029248.1| Mouse hepatitis virus strain MHV-A59 C12 mutant, complete genome  
TATAAGAGTGATTGGCGTCCGTACGTACCCTCTCAACTCTAAAACCTCTTGTAGTTTAAATCTAATCTAAA  
CTTTATAAACGGCACTTCCTGCGTGTCCATGCCGCGGGCCTGGTCTTGTTCATAGTGCTGACATTTGTAG  
TTCCTTGACTTTCGTTCTCTGCCAGTGACGTGTCCATTCGGCGCCAGCAGCCCACCCATAGGTTGCATAA  
TGGCAAAGATGGGCAAATACGGTCTCGGCTTCAAATGGGCCCCAGAATTTCCATGGATGCTTCCGAACGC  
ATCGGAGAAGTTGGGTAACCCTGAGAGGTCAGAGGAGGATGGGTTTTGCCCTCTGCTGCGCAAGAACCG  
AAAGTTAAAGGAAAACTTTGGTTAATCACGTGAGGGTGAATTGTAGCCGGCTTCCAGCTTTGGAATGCT  
GTGTTCACTGCTGCCATAATCCGTGATATTTTTGTAGATGAGGATCCCCAGAAGGTGGAGGCCTCAACTAT  
GATGGCATTGCAGTTCGGTAGTGCCGTCTTGGTTAAGCCATCCAAGCGCTTGTCTATTCAGGCATGGACT  
AATTTGGGTGTGCTTCCCAAACAGCTGCCATGGGGTTGTTCAAAGCGCTCTGCCTGTGTAACACCAGGG  
AGTGCTCTTGTGACGCCACGTGGCCTTTCACCTTTTTACGGTCCAACCCGATGGTGTATGCCTGGGTAA  
TGGCCGTTTTATAGGCTGGTTCGTTCCAGTCACAGCCATACCGGAGTATGCGAAGCAGTGGTTGCAACC  
TGGTCCATCCTTCTTCGTAAGGGTGGTAACAAAGGGTCTGTGACATCCGGCCACTTCCGCCGCGCTGTTA  
CCATGCCTGTGTATGACTTTAATGTAGAGGATGCTTGTGAGGAGGTTCACTTAAACCCGAAGGGTAAAGTA  
CTCCTGCAAGGCGTATGCTCTTCTTAAGGGCTATCGCGGTGTTAAGCCATCCTGTTTGTGGACCAGTAT
```

Choose the sequences

- DNA or protein sequence: far related can use protein sequence
 - Nucleotide
 - Amino acid
 - nature selection act on protein.
 - different translation table: species, organelle

	{	Ile (N)
AUA		Met (M)
AUG		Met

Start with POWER



The Phylogenetic Web Repeater (POWER) allows users performing phylogenetic analysis with molecular data by most programs of PHYLIP package repeatedly. POWER provide two pipelines to process the analysis. One of them includes multiple sequence alignment (MSA) at the beginning of the pipeline whereas the other begin phylogenetic analysis with aligned sequence.

Please start your analysis by selecting the pipeline and the data type:

Pipeline	<ul style="list-style-type: none">◦ MSA + Phylogenetic Analysis(Input the FASTA format)◦ Phylogenetic Analysis Only(Input the PHYLIP format)
Sequence Type	<ul style="list-style-type: none">◦ DNA◦ Protein



Alignment program-ClustalX (for Windows)

ClustalX (1.81)

File Edit Alignment Trees Colors Quality Help

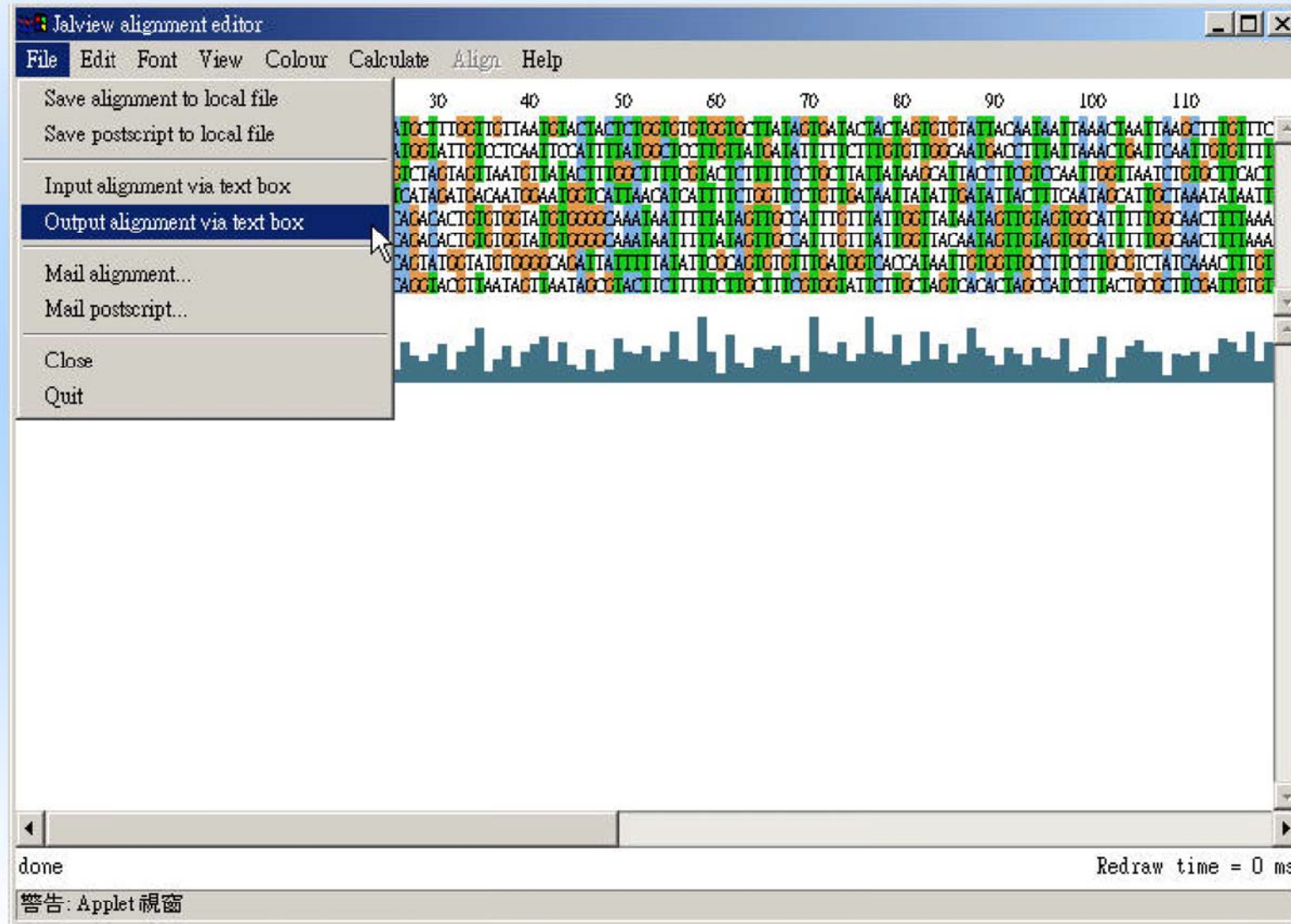
Multiple Alignment Mode Font Size: 10

Species	Sequence
1 Volpe	MKGS ^{ED} LKKHGNTVLTALGGILKKKG ^H HEAELKFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^D
2 Cane	MKGS ^{ED} LKKHGNTVLTALGGILKKKG ^H HEAELKFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^D
3 Uomo	MKASE ^D LKKHGATVLTALGGILKKKG ^H HEAEIKFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^E
4 Scimpanzè	MKASE ^D LKKHGATVLTALGGILKKKG ^H HEAEIKFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^E
5 Gorilla	MKASE ^D LKKHGATVLTALGGILKKKG ^H HEAEIKFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^E
6 Gibbone	MKASE ^D LKKHGATVLTALGGILKKKG ^H HEAEIKFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^E
7 Orango	MKASE ^D LKKHGATVLTALGGILKKKG ^H HEAEIKFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^E
8 Macaco	MKASE ^D LKKHGVTVLTALGGILKKKG ^H HEAEIKFLA ^Q SHA ^T KKHKIPV ^K YLELIS ^E
9 Pipistrello	MKASE ^D LKKHGATVLTALGGILKKKG ^Q HEAQLKFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^E
10 Maiale	MKASE ^D LKKHGNTVLTALGGILKKKG ^H HEAELTFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^E
11 Cavallo	MKASE ^D LKKHGTWVLTALGGILKKKG ^H HEAELKFLA ^Q SHA ^T KKHKIPV ^K YLEFIS ^D
12 Pollo	MKGS ^{ED} LKKHGATVLTQLGKILKQKGNH ^E SELKFLA ^Q THATKKHKIPV ^K YLEFIS ^E

ruler 60 70 80 90 100 110

CLUSTAL-Alignment file created []

Jalview for Repeated Job (in POWER)



The screenshot displays the Jalview alignment editor window. The title bar reads "Jalview alignment editor". The menu bar includes "File", "Edit", "Font", "View", "Colour", "Calculate", "Align", and "Help". The "File" menu is open, showing options: "Save alignment to local file", "Save postscript to local file", "Input alignment via text box", "Output alignment via text box" (highlighted), "Mail alignment...", "Mail postscript...", "Close", and "Quit".

The main window shows a sequence alignment with a color-coded bar above it. The alignment is displayed in a grid format with columns numbered 30, 40, 50, 60, 70, 80, 90, 100, and 110. The alignment consists of several lines of nucleotide sequences (A, C, G, T) with gaps (represented by dashes). Below the alignment is a bar chart showing the conservation of each position across the sequences.

At the bottom of the window, the status bar displays "done" on the left and "Redraw time = 0 ms" on the right. A warning message in Japanese is visible at the very bottom: "警告: Applet 視窗".

Phylogenetic Analysis Step 2

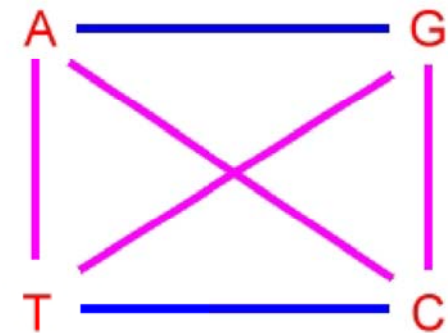


- Sequences alignment
 - ClustalW, Jalview
- Build substitution model
 - Parameters determined



Substitution model

- Transition: purine to purine, pyrimidine to pyrimidine
- Transversion: purine to pyrimidine (and pyrimidine to purine)
- Base frequencies
- Among-site rate variation: Wobble hypothesis
 - Invariable, Variable sites
- Outgroup: to root a tree



— Transition
— Transversion

Substitution model of Distance methods



Jukes-Cantor

Substitution model

Jukes-Cantor

Kimura 2
parameter

Substitution model

Kimura 2 parameter

Transition/transversion ratio

2 (must be a positive real number)

Jin and Nei

Substitution model

Jin and Nei

Transition/transversion ratio

2 (must be a positive real number)

Coefficient of variation of
substitution rate among site

(must be a positive real number)

ML

Substitution model

Maximum Likelihood

Transition/transversion ratio


2 (must be a positive real number)

Use empirical base frequencies

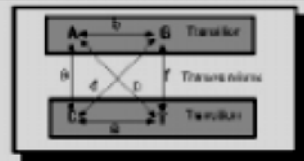
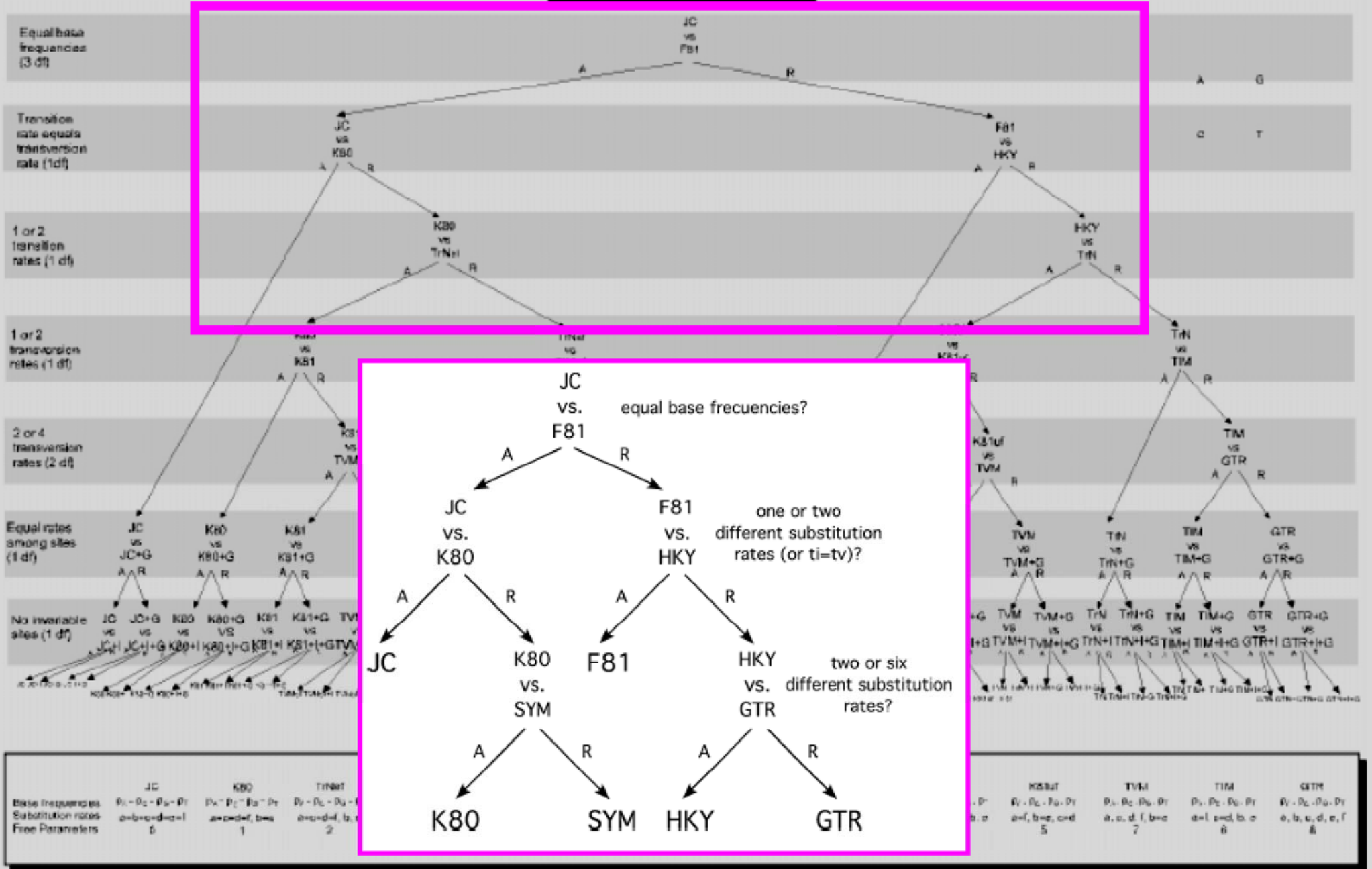
Yes No, entry base frequencies for A, C, G, T/U

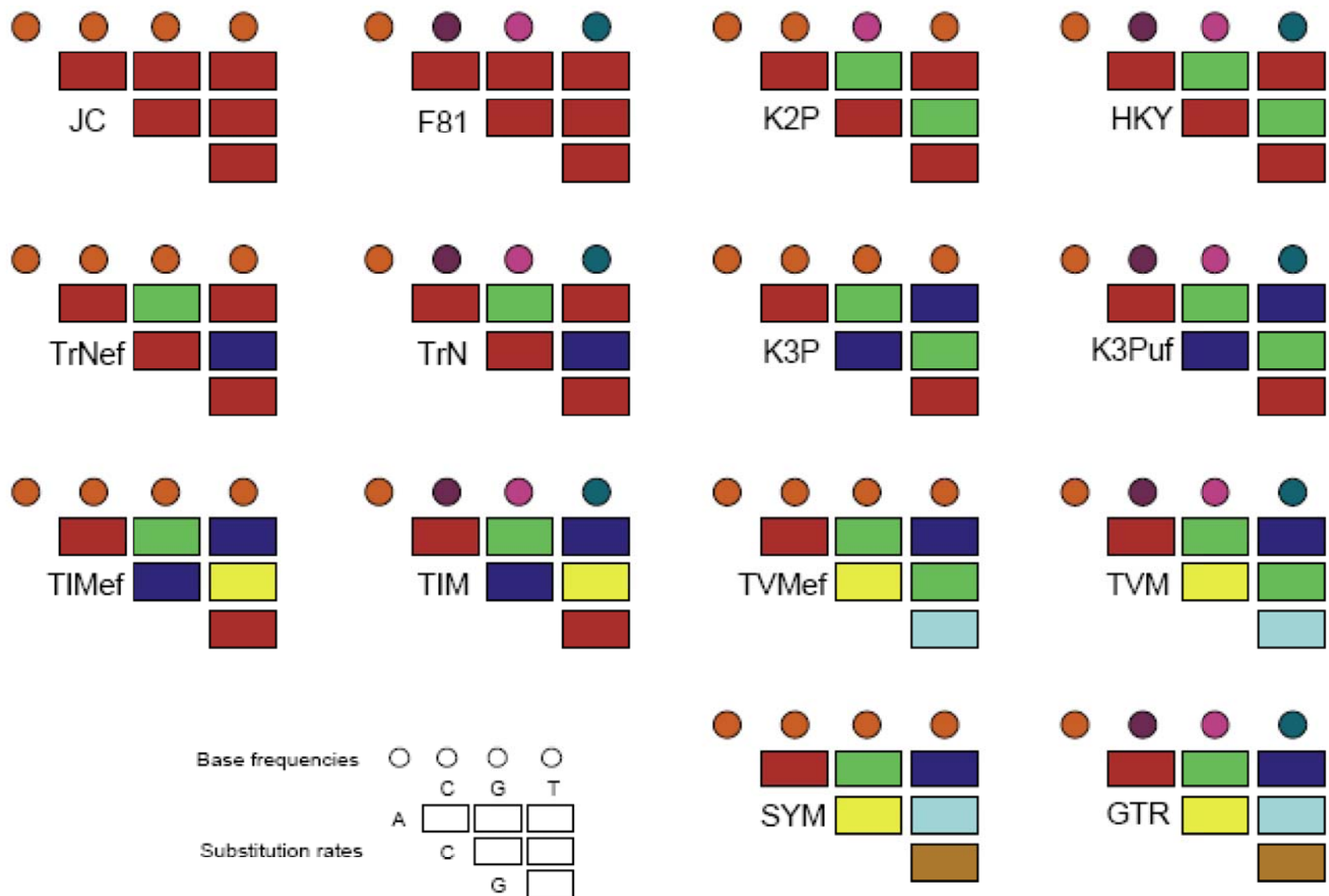


Choose the suitable model : Model selection

- Which method would be better for our data matrix?
- Using a variety of method to analysis the data matrix
- Software: Modeltest 3.7 
 - <http://darwin.uvigo.es/software/modeltest.html>

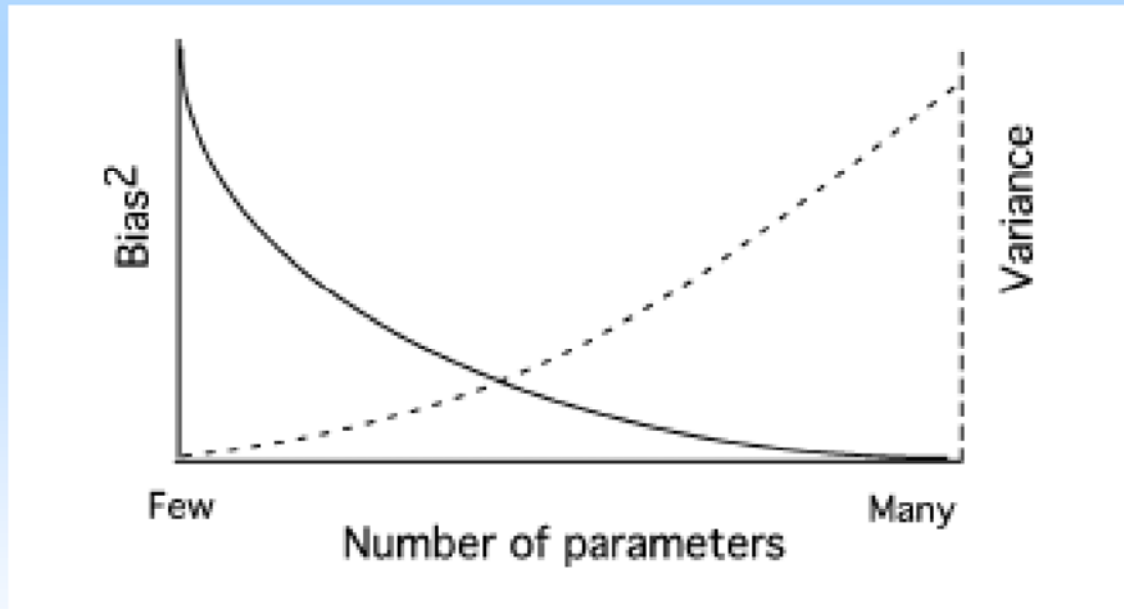
Modeltest 3.0 hierarchy





Parameter in the model

- More parameters the model uses, more powerful it does?



Bias: distance between the average estimate and truth

Variance: spread of the estimates around the truth

Phylogenetic Analysis Step 3



- Sequences alignment
 - ClustalW, Jalview
- Build substitution model
 - Parameters determined
- Construct phylogenetic tree
 - Quantitatively determine the relationship between
input sequences



Phylogenetic Analysis Step 3



- Sequences alignment
 - ClustalW, Jalview
- Build substitution model
 - Parameters determined
- Construct phylogenetic tree
 - Quantitatively determine the relationship between input sequences
- Evaluate phylogenetic tree

Pylogenetic analysis for DNA

Character state methods

- [Maximum parsimony\(heuristic szech\) method](#)
- [Maximum parsimony\(branch and bound search\) method](#)
- [Compatibility method](#)

Distance methods

- [Neighbor-joining and UPGMA method](#)
- [Fitch-Margoliash and least squares method](#)
- [Fitch-Margoliash and least squares method with molecular clock](#)

Maximum likelihood methods

- [Maximum likelihood method](#)
- [Maximum likelihood method with molecular clock](#)

(Warning: these methods cost much memory. So, if the number of bases of your sequences data exceeds 40000, the job may fail.

Pylogenetic analysis for Protein



Character state methods

- Maximum parsimony(heuristic search) method

Distance methods

- Neighbor-joining and UPGMA method
- Fitch-Margoliash and least squares method
- Fitch-Margoliash and least squares method with molecular clock

Phylogenetic Analysis Step 4



- Sequences alignment
 - ClustalW, Jalview
- Build substitution model
 - Parameters determined
- Construct phylogenetic tree
 - Quantitatively determine the relationship between input sequences
- Evaluate phylogenetic tree



Evaluate reliability of tree

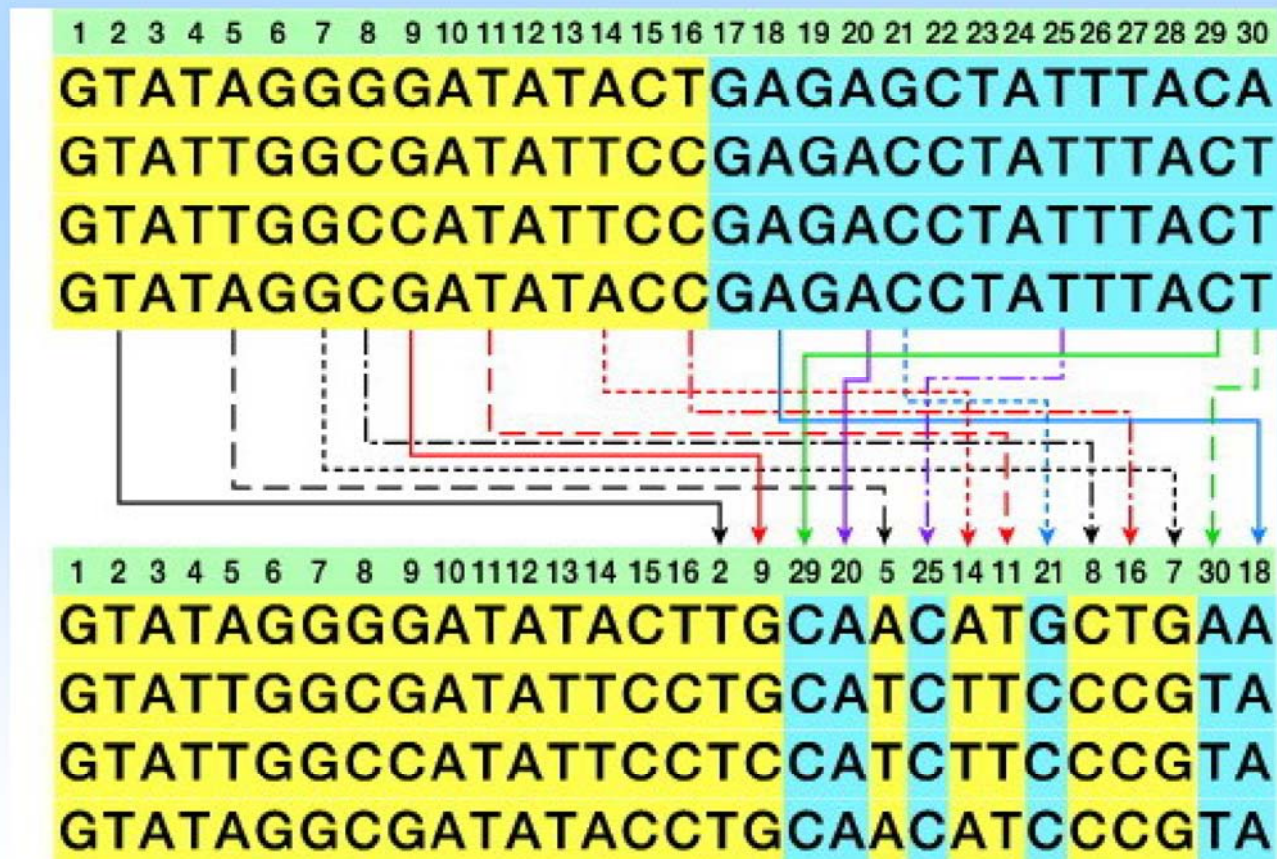


- Sequences resampling
 - Bootstrap
 - Delete-half jackknife
 - Permute species for each character

- Get the same results with different methods

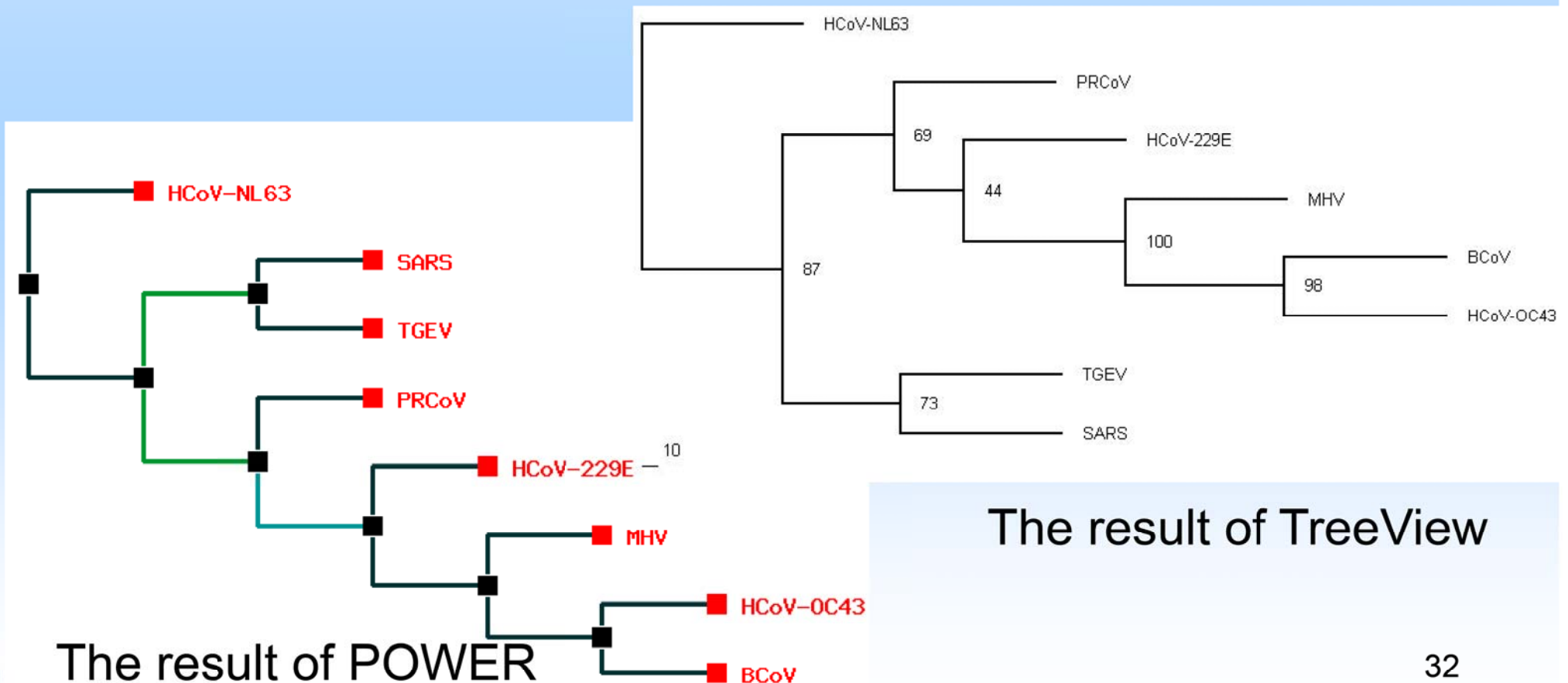
Bootstrapping

- Generate pseudo-sequence alignment randomly
- Compare with original result to get bootstrap value

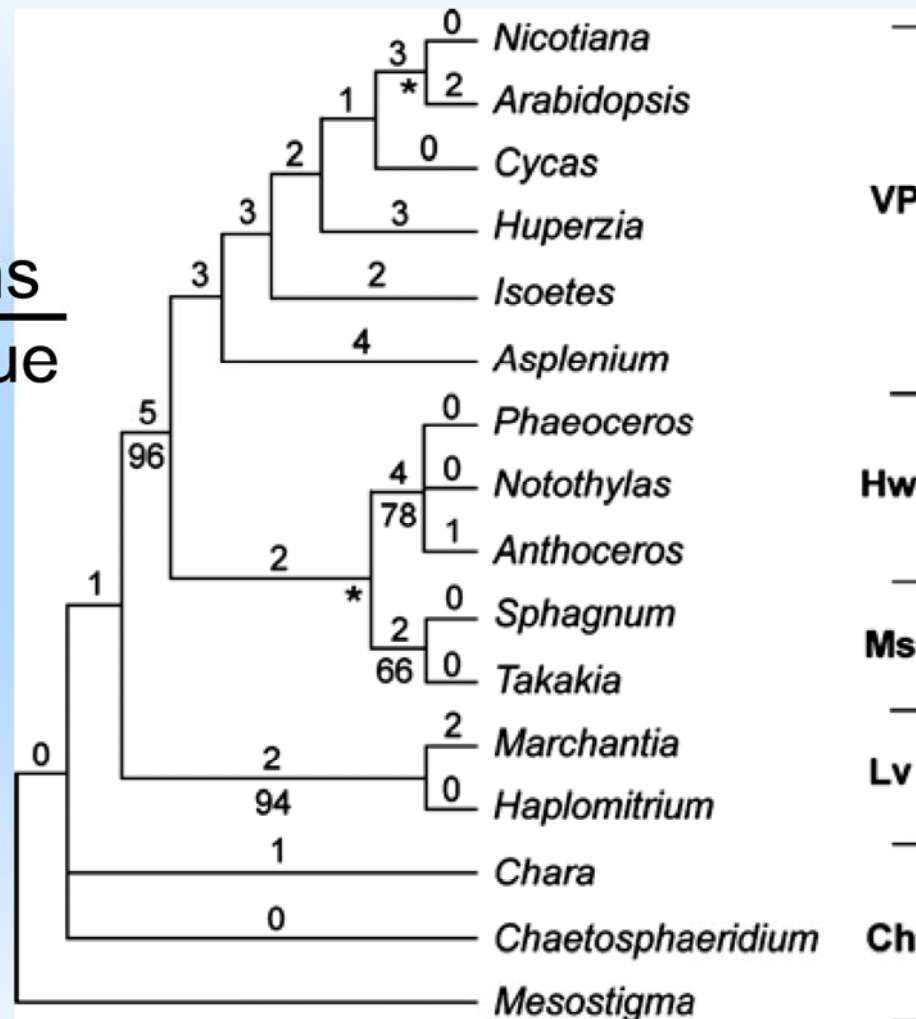


Construct trees with Bootstrapping

Reliability measurement: The probability that the nodes of a given clade are always nodes of that clade

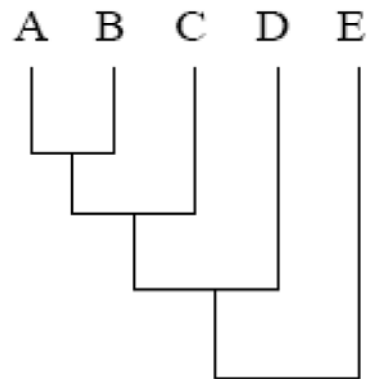


Branch lengths
Bootstrap value

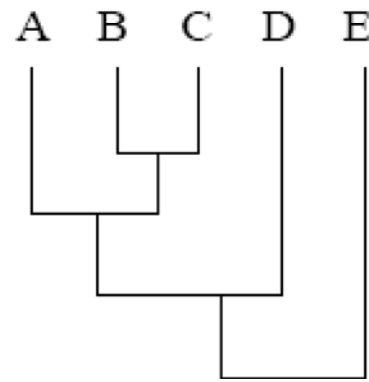


(Yin-Long Qiu et al. 2006 PNAS)

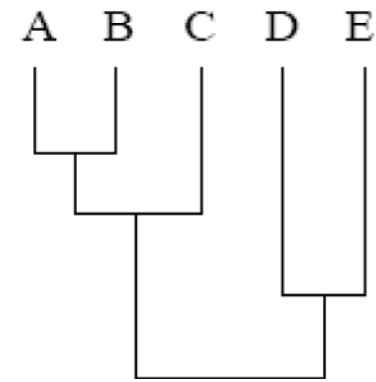
Consensus trees



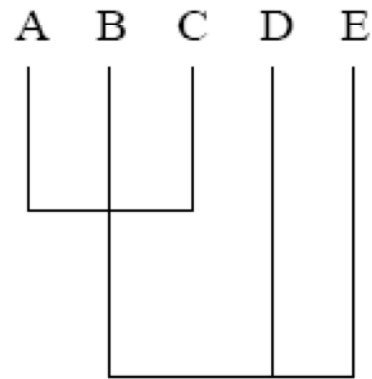
Tree 1



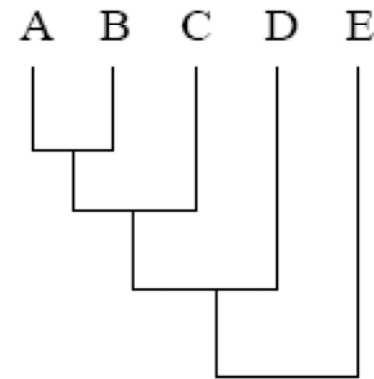
Tree 2



Tree 3



Strict
consensus tree



Majority-rule
consensus tree

PhylOgenetic Web Repeater (POWER)

Result and Logs

Online or as bookmark



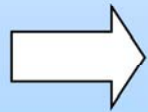
Dear Sir,

We accepted your submission. The job will be done in a few minutes to hours. After job finished, you will receive a notice email. Or You can check the result from the link below.

http://power.nhri.org.tw/power/result_page.php?job_no=28598&job_name=my_job_0215_090002

Thanks for using POWER. Any comment will be appreciated.

Your faithfully,
POWER Administrator.



Or E-mail notification



Subject: [POWER]Job 'comonavirus0720' Finished at 2004-07-20 18:06:36

Dear Sir or Madam:

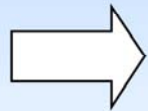
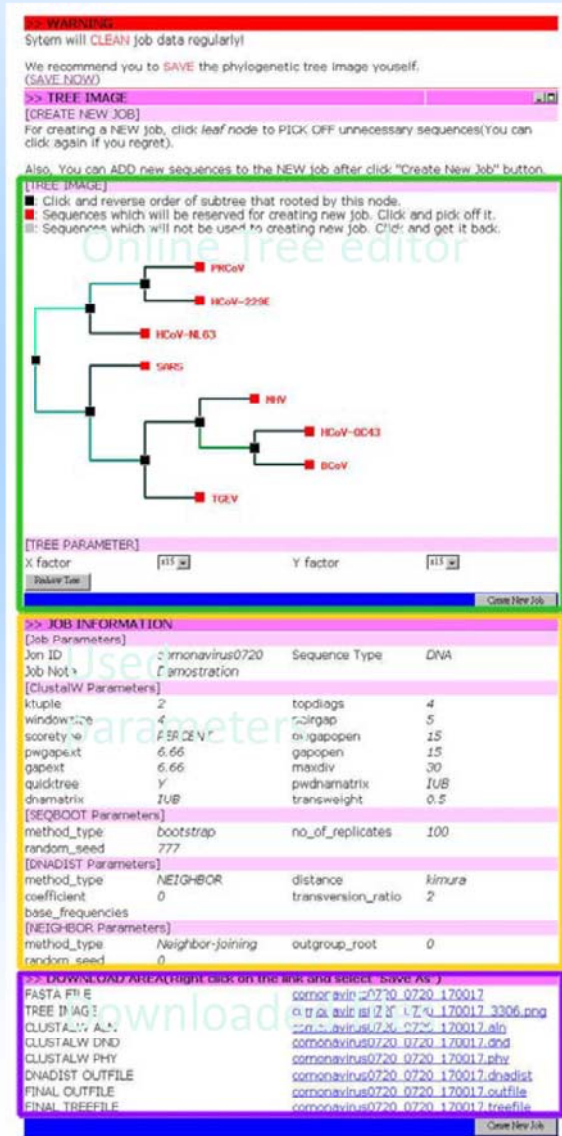
The job 'comonavirus0720' you sent at 2004-07-20 16:00:37 has finished!
The whole process that started at 2004-07-20 18:06:13 and finished at 2004-07-20 18:06:36 cost 00:00:22.
You can check the result from the link below.
Thank you for using POWER.

Your faithfully,
POWER Administrator

Job ID: comonavirus0720
Job Note:
Download:
http://211.76.166.77/power/result_page.php?job_no=2841&job_name=comonavirus0720_0720_120017

POWER version 1.0
PHYML package version 3.5
ClustalW version 1.82

May the POWER with you.

System will CLEAN job data regularly!

We recommend you to SAVE the phylogenetic tree image yourself.
(SAVE NOW)

>> TREE IMAGE

[CREATE NEW JOB]

For creating a NEW job, click: leaf node to PICK OFF unnecessary sequences (You can click again if you regret).

Also, You can ADD new sequences to the NEW job after click: "Create New Job" button.

[TREE IMAGE]

- Click and reverse order of subtree that rooted by this node.
- Sequences which will be reserved for creating new job. Click and pick off it.
- Sequences which will not be used to creating new job. Click and get it back.

Online tree editor

[TREE PARAMETER]

X factor [115] Y factor [115]

>> JOB INFORMATION

[Job Parameters]

Job ID	comonavirus0720	Sequence Type	DNA
Job Note	demonstration		

[ClustalW Parameters]

ktuple	2	topdiags	4
windowsize	4	scaling	5
scoretype	FASTDEV	onigapopen	15
pwgapext	6.66	gapopen	15
gapext	6.66	maxdiv	30
quidtree	Y	pwdnamatrix	IUB
dnamatrix	IUB	transweight	0.5

[SEQBOOT Parameters]

method_type	bootstrap	no_of_replicates	100
random_seed	777		

[DNADIST Parameters]

method_type	NEIGHBOR	distance	kimura
coefficient	0	transversion_ratio	2

base_frequencies

[NEIGHBOR Parameters]

method_type	Neighbor-joining	outgroup_root	0
random_seed	0		

>> DOWNLOAD AREA (right click on the link and select: "Save As...")

FASTA FILE comonavirus0720_0720_120017.comonavirus0720_0720_120017.fasta

TREE IMAGE comonavirus0720_0720_120017.tree.png

CLUSTALW ALI comonavirus0720_0720_120017.ali

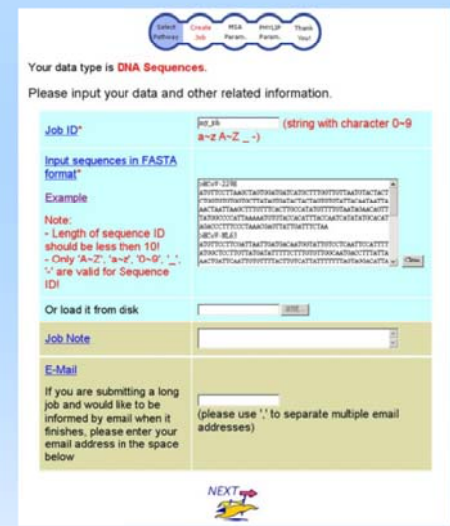
CLUSTALW DND comonavirus0720_0720_120017.dnd

CLUSTALW PHY comonavirus0720_0720_120017.phy

DNADIST OUTFILE comonavirus0720_0720_120017.dnadist

FINAL OUTFILE comonavirus0720_0720_120017.outfile

FINAL TREEFILE comonavirus0720_0720_120017.treefile

Your data type is DNA Sequences.

Please input your data and other related information.

Job ID* [my_job] (string with character 0-9 a-z A-Z _)

Input sequences in FASTA format

Example

Note:

- Length of sequence ID should be less than 10!
- Only 'A-Z', 'a-z', '0-9', '_', '.' are valid for Sequence ID!

Or load it from disk [Browse]

Job Note []

E-Mail

If you are submitting a long job and would like to be informed by email when it finishes, please enter your email address in the space below []

Next

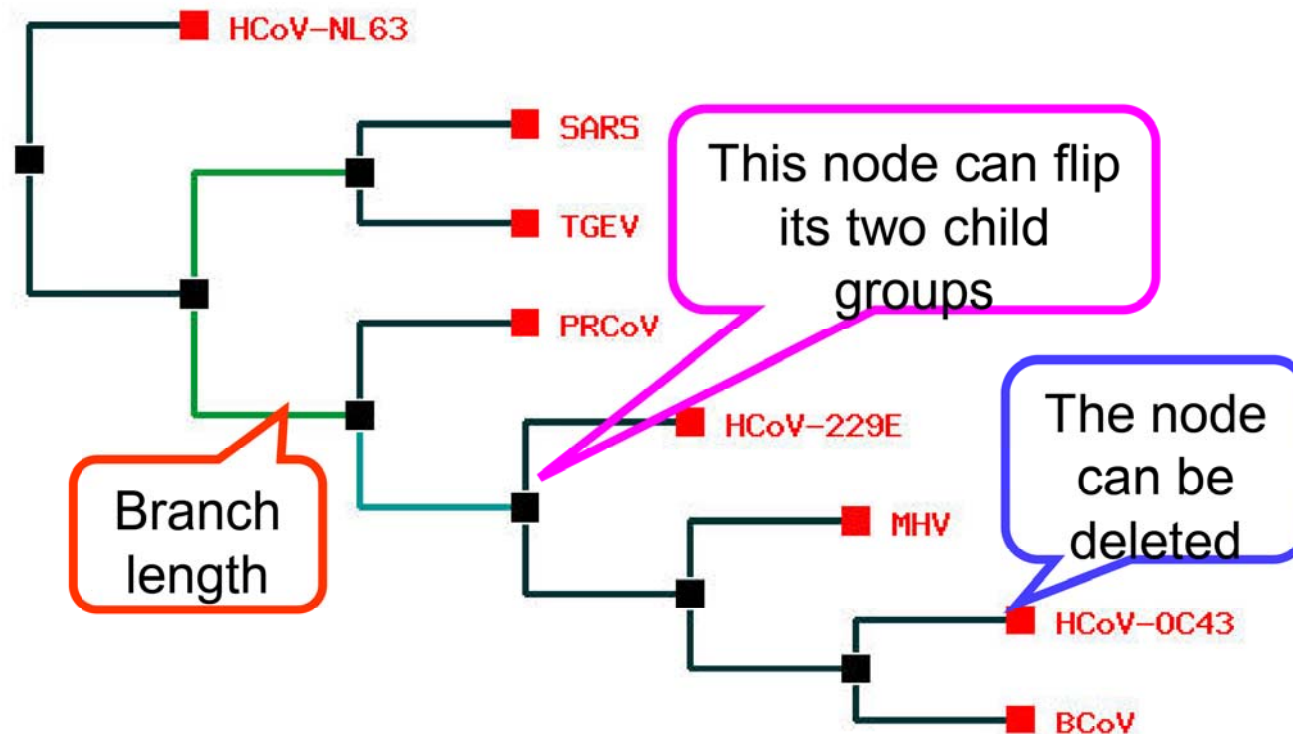
Re-perform the process by items added or deleted



The Result of POWER

Color of line indicates how many times the group which consists of the species to the right of the fork occurred.

—: ≤ 20 —: >20 & ≤ 40 —: >40 & ≤ 60 —: >60 & ≤ 80 —: >80



The files can downloaded



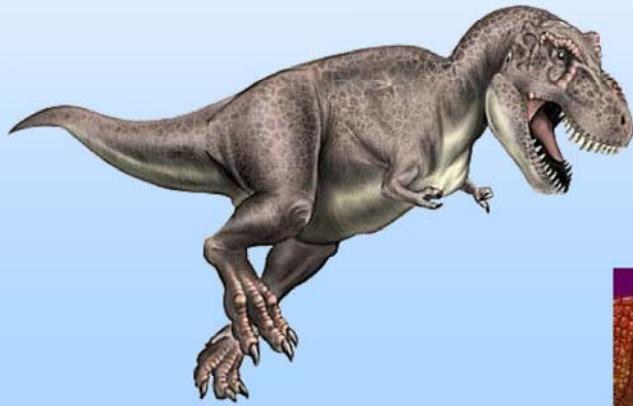
- Original fasta file
- .phy (generated by clustalw)
- Tree Image
- outfile(generated by phylip programs)
 - Consensus file if bootstrap had be made
- treefile

Reconstruct tree completely??



- Species evolution is a history, which can't be reconstructed fully
- All models are questionable but some are useful
- Narrow down difference between gene tree and species tree

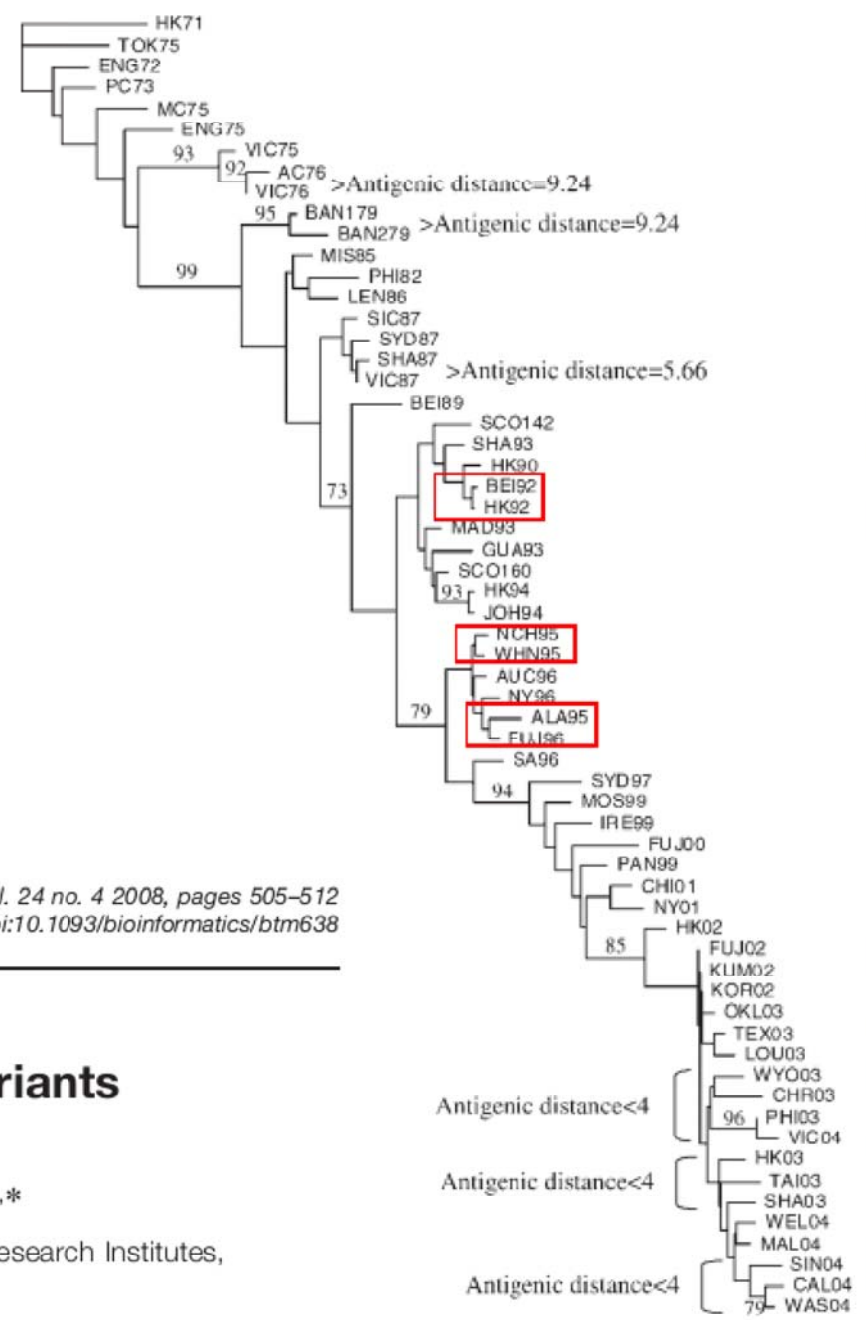
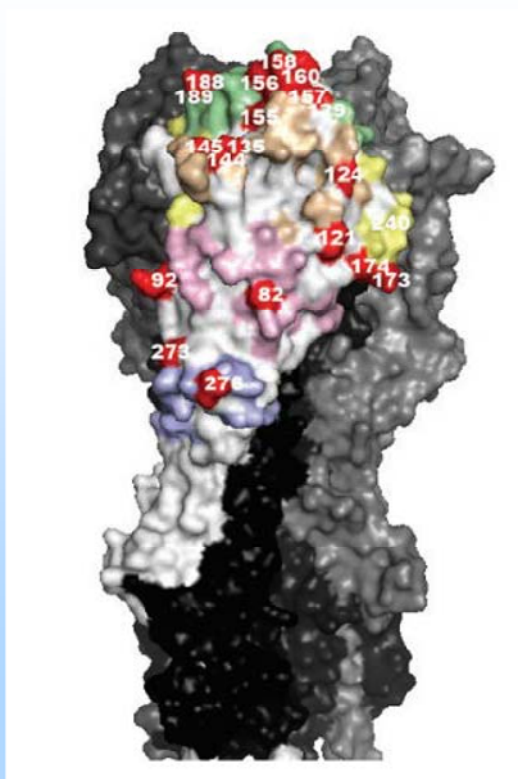
Tyrannosaurus



Reconstruct tree completely??



- Species evolution is a history, which can't be reconstructed fully
- All models are questionable but some are useful
- Narrow down difference between gene tree and species tree



Sequence analysis

Bioinformatics models for predicting antigenic variants of influenza A/H3N2 virus

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Fig. 3. Neighbor-joining phylogenetic tree of the 62 influenza H3N2 viruses (HA1 protein with 329 amino acid residues). The full names of viruses are in the Supplementary Data. Numbers at branch nodes refer to the percentage of 1000 bootstrap repetitions, and only those occurring at a percentage >70% are shown.

NJ with 1000 Replicates in POWER

The Journal of
**Experimental
Biology**

The Journal of Experimental Biology 212, 1202-1211
Published by The Company of Biologists 2009
doi:10.1242/jeb.025239

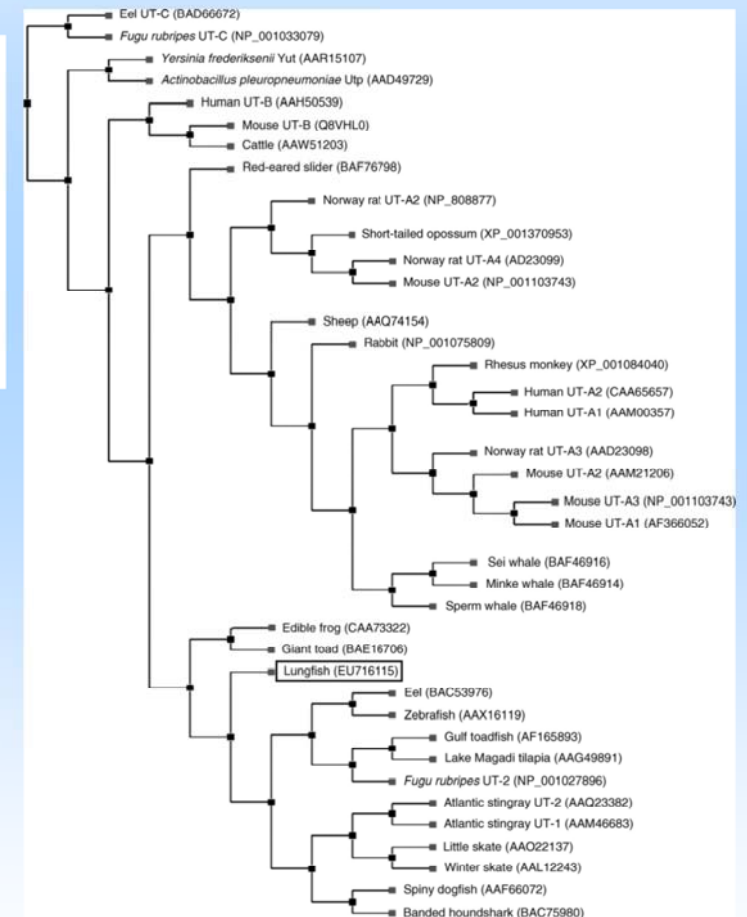
Increased gene expression of a facilitated diffusion urea transporter in the skin of the African lungfish (*Protopterus annectens*) during massively elevated post-terrestrialization urea excretion

Carrie Y. C. Hung¹, Fernando Galvez², Yuen K. Ip³ and Chris M. Wood^{1,*}

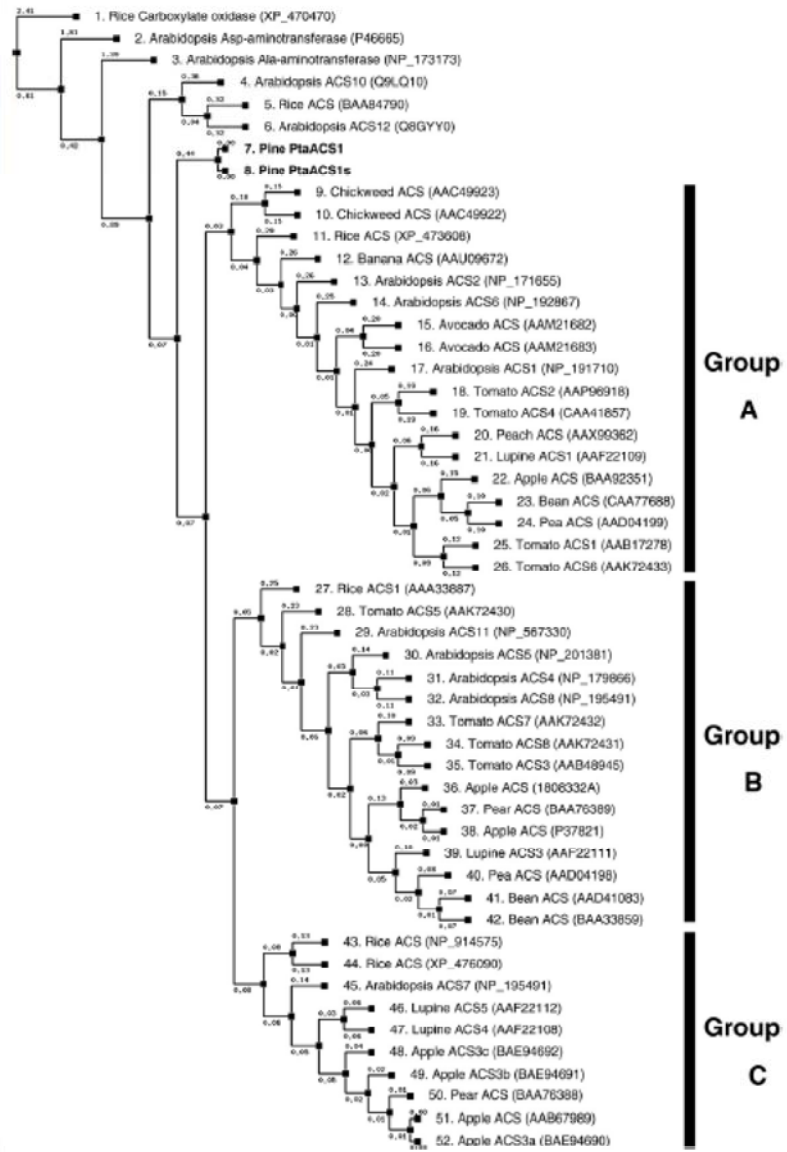
¹Department of Biology, McMaster University, Hamilton, ON, Canada, L8S 4K1, ²Department of Biological Sciences, Louisiana State University, Baton Rouge, LA 70803, USA and ³Department of Biological Sciences, National University of Singapore, 10 Kent Ridge Road, Singapore 117543, Republic of Singapore

Phylogenetic tree of urea transporters across taxa. The protein sequences were aligned using ClustalW software, followed by neighbour-joining (NJ) matrix for tree reconstruction and evaluated by means of a bootstrap of 1000 replicates at

<http://power.nhri.org.tw>



Perform ML in POWER



Available online at www.sciencedirect.com



Gene 413 (2008) 18–31

GENE

www.elsevier.com/locate/gene

Characterization of a 1-aminocyclopropane-1-carboxylate synthase gene from loblolly pine (*Pinus taeda* L.)

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Phylogenetic tree depicting the relatedness of ACC synthase and aminotransferase protein sequences in GenBank. The phylogenetic tree was generated using the **POWER server** (<http://power.nhri.org.tw/>) with default parameters for the maximum likelihood method (ML) and molecular clock, but without bootstrapping or random input of sequences.



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